

Review

# Nothing in Evolution Makes Sense Except in the Light of Genomics: Read–Write Genome Evolution as an Active Biological Process

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**Abstract:** The 21st century genomics-based analysis of evolutionary variation reveals a number of novel features impossible to predict when Dobzhansky and other evolutionary biologists formulated the neo-Darwinian Modern Synthesis in the middle of the last century. These include three distinct realms of cell evolution; symbiogenetic fusions forming eukaryotic cells with multiple genome compartments; horizontal organelle, virus and DNA transfers; functional organization of proteins as systems of interacting domains subject to rapid evolution by exon shuffling and exonization; distributed genome networks integrated by mobile repetitive regulatory signals; and regulation of multicellular development by non-coding lncRNAs containing repetitive sequence components. Rather than single gene traits, all phenotypes involve coordinated activity by multiple interacting cell molecules. Genomes contain abundant and functional repetitive components in addition to the unique coding sequences envisaged in the early days of molecular biology. Combinatorial coding, plus the biochemical abilities cells possess to rearrange DNA molecules, constitute a powerful toolbox for adaptive genome rewriting. That is, cells possess “Read–Write Genomes” they alter by numerous biochemical processes capable of rapidly restructuring cellular DNA molecules. Rather than viewing genome evolution as a series of accidental modifications, we can now study it as a complex biological process of active self-modification.

**Keywords:** genome sequence; molecular phylogeny; repetitive DNA; symbiogenesis; hybrid speciation; whole genome duplication; horizontal DNA transfer; viviparity; mobile DNA elements; stem cell pluripotency

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## 1. Introduction

The title of this mini-review is a paraphrase of Dobzhansky’s famous dictum, “Nothing in biology makes sense except in the light of evolution” [1,2]. The reason for this paraphrase is to emphasize how, since Dobzhansky’s day, genetics and evolution science have moved into the era of a revolutionary new technology, DNA sequencing. Whenever such a technological revolution occurs, science must always ask itself what impact data from the new methods has on the validity of prevailing concepts.

Our current ideas of heredity center on DNA replication and transmission [3]. Although cell and organismal heredity involves transmission of all cellular molecules, we conventionally view RNAs, proteins, lipids, polysaccharides and other biomolecules as derivative products resulting from biochemical activities determined by genomic DNA sequences [4]. According to this DNA-centered perspective, stable inherited changes in organismal properties result primarily from alterations in the genome. New DNA sequences can encode new biochemical capabilities that lead to novel traits. If evolution is the acquisition of new characters over time [5], the most basic causal events should emerge from the processes that generate new DNA sequences. Those events are traceable in genome sequences, which today serve as the ultimate empirical data to test evolutionary hypotheses.

In this brief review, we will examine some of the evolutionary lessons from genome sequence data. Given the broad extent of the subject matter, only a select range of examples will receive attention. The investigation shows that many evolutionary changes result from cellular processes that produce abrupt changes in genome structure and organismal characters. These processes include symbiogenesis, horizontal DNA transfer, hybrid speciation and natural genetic engineering, especially the action of mobile DNA elements [6,7]. At the molecular level, evolution is often saltational rather than gradual [8], and punctuated equilibrium [9] is the default pattern to expect in the history of phenotypic properties. As we shall see, this view is quite different from Dobzhansky's thinking. In particular, molecular sequence analysis documents many genomic elements and novel forms of hereditary variation that were hardly conceivable in Dobzhansky's lifetime.

## 2. The Genome as a Highly Formatted Sequence Database

Genomes serve as controlled read-write databases for the reliable transmission and regulated expression of DNA sequence information [7,8,10]. Because controls over expression, replication and transmission are essential to survival and reproduction, genomes contain abundant *cis*-acting DNA formatting elements in addition to coding sequences determining RNA and protein primary structures [11].

Many of the formatting DNA elements are generic and present at multiple genome locations. Consequently, repetitive DNA is a major component of all genomes, and repeats frequently represent the majority of an organism's DNA [12]. In the human genome, the repetitive content is about two-thirds of the total DNA, compared to well under 5% for protein-coding sequences [13].

For evolution, the significance of genome formatting by repeat DNA elements is that important phenotypic alterations can occur as a result of changes to so-called "non-coding" sequences. Recent discoveries about the functions of "non-coding" ncRNA molecules illustrate this point [14–19], as do Evo-Devo findings that many developmental alterations distinguishing related organisms occur in regulatory DNA elements rather than coding sequences [20,21].

## 3. Molecular Phylogenies Based on Core Information-Processing Systems

The most basic impact of genomics on evolution science has been the use of sequence data to establish relationships among different organisms. In the 1970s, Carl Woese pioneered molecular phylogenetic methodologies [22]. Woese chose to base his initial phylogenies on the sequence of small subunit ribosomal RNA for two reasons: (i) ssrRNA was abundant and amenable to 1970s sequence analysis methods; and (ii) the ribosome is a highly conserved central component of information transfer from the genome to the proteome. Because all cells have similar but still distinctive ribosomes, this organelle provided phylogenetic data to establish connections between very diverse organisms.

Using ssrRNA sequence analysis, Woese and his colleagues unexpectedly discovered that there are in fact two separate kingdoms of prokaryotic organisms (those lacking separate cell nuclei), not one bacterial kingdom as previously believed [23]. The two groups of prokaryotic organisms turned out to exhibit ssrRNA sequence clusters as phylogenetically distant from each other as both clusters were from the ssrRNA sequences of eukaryotic (nucleated) organisms. The evolutionary distance between the two prokaryotic groups was further confirmed by major differences in their cell membranes as well as by differences in the basic processes of genome replication and expression. The newly discovered prokaryotic cell kingdom was labeled Archaea because the first members to be studied were organisms isolated from extreme environments thought to resemble the early Earth [24]. Today, however, there is no reason to assume that Archaea are any more ancient than Bacteria.

The most basic tenet of evolution science—the genetic relatedness of all living organisms—is abundantly supported by molecular biology, in particular the universal features of the triplet code for amino acids and the similarities of core cell structures, like the ribosome, associated translation factors [25], and DNA and RNA polymerases. Nonetheless, the first phylogenetic application of genomics revealed a previously unknown complexity in the biosphere and, as we shall see in the

next section, provided compelling evidence for a large number of evolutionary processes excluded by conventional evolutionary thinking based on population genetics and the Modern Synthesis [26,27]. As highlighted below, the phylogenies of Bacteria, Archaea, eukaryotes and giant viruses continue to pose intriguing challenges in understanding the networked evolutionary connections between all domains of life [28].

#### 4. Eukaryotic Origins and Major Eukaryotic Taxonomic Originations through Symbiogenesis

The identification of an unsuspected bifurcation among prokaryotes, the most abundant living organisms [29], immediately raised questions about the historical relationships between Archaea and Bacteria without nuclei, on the one hand, and Eukarya, the organisms with nucleated cells, on the other. Eukaryotes formed a coherent separate group based on ribosomal RNA sequences, and molecular phylogenies of different eukaryotic groups confirmed well-established taxonomic classifications, such as fungi, plants and animals. But the generic eukaryotic cell was closer to Bacteria in some features—membrane composition, metabolic pathways—and closer to Archaea in other features—replication, transcription and translation [30,31]. This phenotypic dichotomy gave support for longstanding but hotly disputed arguments championed by Lynn Margulis and others that symbiogenetic cell fusions served to create complex eukaryotic cells from simpler prokaryotic progenitors [29,32–36].

##### 4.1. Endosymbiotic Bacterial Origins of Eukaryotic Organelles

Molecular phylogenetics unequivocally documented the role of symbiogenesis in Eukarya evolution by analysis of the two cell organelles that have their own genomes: (1) the mitochondrion, carrying out oxidative metabolism; and (2) the chloroplast, carrying out oxidative photosynthesis. All eukaryotic cells contain a functional mitochondrion or a non-oxidative derivative organelle [37–39]. Thus, the ancestral eukaryote must have acquired an endosymbiotic mitochondrial precursor, and molecular phylogenetics unambiguously identified the mitochondrion as belonging to the Alpha-proteobacteria group [40–42]. Similarly, photosynthetic eukaryotes, including red and green algae and plants, have chloroplasts, and molecular phylogenetics identified the chloroplast as a member of the cyanobacteria [43–45]. The chloroplast, and derivative plastid organelles in phylogenetically related non-photosynthetic eukaryotes, therefore must have descended from one or more endosymbiotic cyanobacteria [46,47].

##### 4.2. DNA Transfer between Endosymbiotic Organelle and Nuclear Genomes

The genomes of mitochondria and chloroplasts do not encode all the proteins inherited from their bacterial ancestors. Many organelle proteins are encoded in the nuclear genome as a result of DNA transfers into the nuclear genome from mitochondria [48–51] and from chloroplasts [52–55]. These intracellular DNA transfers from one genomic compartment to another continue and have been analyzed in real time [56–60]. Interestingly, mitochondrial DNA transfer to the nucleus affects life-span in yeast cells [61], increases with age in rat cells [62], generates inherited disease loci in humans [56,63], occurs frequently in cancer cells [64], and participates in nuclear double-strand DNA break repair in organisms as distant as yeast and humans [56,65]. Chloroplast to nucleus DNA transfer is stimulated by stress conditions [59]. There is further evidence of organelle genomes acquiring nuclear and other external DNA [66–68]. Active import of DNA into plant mitochondria has been documented in real time [69].

Since the genetic content of mitochondria, chloroplasts and plastids differs significantly among taxonomic groups, important factors in eukaryotic taxonomic divergence are organelle to nucleus DNA transfer [49,50,70] and organelle genome restructuring: in mitochondria [51,71–87], by loss of mitochondrial oxidative capacity [38,88–90], in chloroplasts [45,47,54,91–96], and by loss of chloroplast photosynthetic capacity [47,54,97–100].

#### 4.3. Origins of Photosynthetic Eukaryotic Taxa by Secondary Endosymbiogenesis

A large number of photosynthetic eukaryotes did not evolve directly from algae or plants and are most closely related taxonomically to non-photosynthetic organisms [37]. These organisms originated from “secondary” eukaryote to eukaryote symbiogenetic events where a red or green alga has become an endosymbiont in the initially non-photosynthetic lineage [101–106]. The resulting photosynthetic cells have four different genome compartments that exchange DNA segments: nucleus, mitochondrion, plastid and nucleomorph (descended from the algal nucleus) [107–113]. As with mitochondria and chloroplasts, the major intracellular DNA transfers occur from the organelles, including the nucleomorph, into the nuclear genome. The photosynthetic taxa arising from secondary endosymbiosis include euglenids and chlorachniophytes from green algal endosymbiosis and the chromalveolates from red algal endosymbiosis [37]. The large chromalveolate phylum includes major photosynthetic organisms responsible for a large fraction of atmospheric oxygen, such as brown algae, dinoflagellates and diatoms.

Photosynthetic endosymbiosis is not restricted to unicellular eukaryotes. There are cases where animals have acquired photosynthetic capabilities by symbiogenetic events [114–116]. The photosynthetic animals include sea slugs [117–119] and molluscs [120,121].

#### 4.4. Formation of a Primitive Eye-Like Organ in a Unicellular Eukaryote by Serial Endosymbioses

Among the photosynthetic dinoflagellates resulting from algal ensymbiosis, there is a group labeled “ocelloids,” which possess a remarkable light-harvesting organ (the ocelloid) that resembles a complex camera-like animal eye [122,123]. The ocelloid has analogues to the cornea, lens, iris and retina. A noteworthy recent paper reports that genomic analysis reveals that each of these structures resulted from a distinct endosymbiogenetic event [124]:

“Here we show, using a combination of electron microscopy, tomography, isolated-organelle genomics, and single-cell genomics, that ocelloids are built from pre-existing organelles, including a cornea-like layer made of mitochondria and a retinal body made of anastomosing plastids. We find that the retinal body forms the central core of a network of peridinin-type plastids, which in dinoflagellates and their relatives originated through an ancient endosymbiosis with a red alga. As such, the ocelloid is a chimaeric structure, incorporating organelles with different endosymbiotic histories.”

The genomics-verified example of ocelloid formation by serial endosymbiogenesis has to be considered in light of Darwin’s famous statement: “If it could be demonstrated that any complex organ existed, which could not possibly have been formed by numerous, successive, slight modifications, my theory would absolutely break down.” [5] (p. 189). While the endosymbiogenetic origin of the ocelloid does not demonstrate the impossibility of camera eyes evolving by Darwinian gradualism, this striking case of adaptive evolution by repeated cell fusion events clearly lies outside the parameters of evolutionary processes envisioned by the author of *Origin of Species* as well as by the Modern Synthesis.

### 5. Eukaryotic Speciation by Inter-Specific Hybridization, Whole Genome Duplications and Genome Restructuring

When we ask how novel species arise from human intervention, it is significant that there are no cases where selection has led to species formation. Selection only modifies existing characteristics by reducing or amplifying them. Artificial species arise through hybridization, as in the case of the wheat-rye hybrid *Triticale* [125,126], and involve genome mergers and whole genome duplication (WGD) events [127,128]. A similar “cataclysmic evolution” process involving hybridization of wild grasses was at the origin of flour wheat (*Triticum*) several thousand years ago and can be reproduced in real time [129,130]. Ongoing abrupt hybrid speciation has been observed to occur in wild sunflowers [131]. A recent paper reports laboratory formation of a novel tobacco species with a double genome by fusion of tissue culture cells from two different natural *Nicotiana* species [132].

Many genomes contain duplicate copies of extended chromosome regions holding homologous genetic loci in a conserved (“syntenic”) order [133–135]. By documenting the prevalence of numerous syntenic duplications, genomic analysis provides compelling evidence that hybrid speciation and WGD have been common events in the history of evolution in all eukaryotic groups, including yeasts and other fungi [136–139], ciliated protists [140], plants [141,142] and animals, including two successive WGD events at the origins of vertebrates [143–147], followed by further WGD events in bony fishes [148,149].

In addition to WGD, interspecific hybridization leads to episodes of genome instability and restructuring [150–152]. Chromosome rearrangements have long been recognized as major features of speciation and taxonomic divergence [153–155]. Repetitive and mobile DNA elements play special roles in chromosome restructuring [156–159], as exemplified in primate evolution by the recently published gibbon genome [160].

## 6. Adaptations Acquired and Comingled by Horizontal Transfers

The conventional view of evolution maintained by Dobzhansky and his colleagues is that traits are transmitted vertically from progenitors to offspring, with evolutionarily important hereditary changes occurring within each particular line of descent. That is the pattern of “descent with modification” illustrated at the end of *Origin of Species* [5]. According to this perspective, each new adaptation has to evolve within a distinct lineage. In contrast to this conventional view, genomic analysis revealed that the molecular phylogenies of genetic loci encoding certain adaptive functions do not always match the taxonomic histories of the basal host genomes [30]. The genomic evidence indicated that organisms could acquire independently evolved DNA providing adaptive benefits from unrelated organisms by “horizontal” DNA transfer [161,162]. Genomic evidence shows that it often proves more efficient to adapt to a new ecological niche by borrowing functions from distant taxa rather than evolving them internally from the pre-existing genome.

### 6.1. Horizontal Transfer among Prokaryotes

The ability of prokaryotic organisms to exchange advantageous DNA segments is evident from studies of antibiotic resistance starting in the early years of molecular genetics (<http://shapiro.bsd.uchicago.edu/ExtraRefs.AntibioticResistanceAndHorizontalTransfer.shtml>) [163–166]. Horizontal DNA transfer can occur between prokaryotic cells by uptake of DNA released by cells to the environment (transformation), direct cell-to-cell contact (conjugation), or viral infection (transduction) [167]. Sometimes the horizontally acquired DNA constituted an independently replicating molecule in the prokaryotic cell [168], and sometimes the horizontally acquired DNA was integrated into the existing genome, often as extended “genomic islands” encoding many different proteins for complex traits like metabolism, defense and pathogenicity [169–172]. Special site-specific recombination structures called “integrons” exist in the genomes of some bacteria for the serial integration of antibiotic resistance cassettes [173,174] and, in particular species, “super-integrons” have accumulated cassettes encoding up to hundreds of diverse adaptive functions [175–178]. Horizontal transfer occurs across the Bacteria-Archaea divide and can even lead to the formation of novel taxa [179–183].

The prevalence of prokaryotic horizontal DNA exchange gave rise to the idea of a vast super-cellular pan-genome that prokaryotes can sample facultatively by transformation, conjugation and transduction to assemble novel genomes for cell adaptation to particular ecological niches [184–186]. This controversial notion has gained wider credibility in recent years as a result of metagenomic analysis, which has revealed unexpectedly large numbers of known and unknown coding functions present in environmental DNA samples, particularly those encapsidated in virus particles [187–193]. There has even been the suggestion that environmental metagenome data indicate the possible existence of major new cell types [194].



### 6.2. Horizontal DNA Transfer from Prokaryotes and Fungi to Multicellular Eukaryotes

Horizontal DNA transfer is not limited to prokaryotes. Genomic analysis provides abundant examples of eukaryotic adaptations with prokaryotic (and fungal) origins. These include:

- Biochemical pathways [195–199];
- Phytopathogenicity in *Botrytis* fungi [200];
- Ability to live in extreme environments [201];
- Capacity of plant parasitic nematodes to digest cellulose and other phytopolymers [202–208]. We know that this horizontal DNA transfer strategy was used repeatedly because each lineage of plant parasitic nematodes acquired their digestive enzymes from different fungi or bacteria;
- Energy metabolism and defense functions subject to purifying selection in a marine shrimp [209];
- Sequences of unknown but selectively conserved function transferred from marine bacteria to fish after the divergence of teleosts from other vertebrates [210].

Many bacteria live as endosymbionts in animals, and there is abundant evidence that parts or all of endosymbiont genomes have been incorporated into host genomes [211–215]. In *Drosophila ananassae*, for example, more than 2% of the genome comes from *Wolbachia* endosymbionts [216].

### 6.3. Horizontal Transfer from Eukaryotes to Bacteria

Although less widely documented than prokaryote to eukaryote horizontal transfer, the analysis of endosymbiotic and pathogenic bacteria infecting eukaryotic cells has turned up examples where these prokaryotes appear to have integrated eukaryotic host cell sequences into their genomes [217–221]. The restricted taxonomic distribution of the eukaryotic domains among the infectious bacteria indicates recent horizontal acquisition rather than shared vertical ancestry with eukaryotes [217].

Bacteria use the proteins containing typically eukaryotic domains encoded by horizontally acquired DNA sequences as injected “effector” molecules to modulate host cell metabolism and defenses in order to facilitate the infection process [222–229]. The ability of many infectious bacteria to grow in diverse eukaryotic hosts, such as amoebae and mammals [230], apparently plays an important role in the acquisition of eukaryotic domains from one kind of host (e.g., amoebae) and their utilization as invasion functions in another kind of host (e.g., mammals).

### 6.4. Horizontal DNA Transfer among Eukaryotes

In addition to fungi, other eukaryotes can transfer DNA horizontally across taxonomic boundaries [162,231]. This phenomenon is prevalent among unicellular protists [232–234], but various classes of DNA transfer have been documented between multicellular lineages:

- Mitochondrial genomes in flowering plants [235–237];
- Chloroplast genomes in plants [238];
- Mobile DNA elements [239–245];
- Sequences encoding diverse adaptive functions, including glyoxylate cycle enzymes in metazoa [246], photosynthetic carbon cycles in plants [247,248], anti-freeze proteins in fish [249], and mimicry pattern determinants in butterflies [250];
- Miscellaneous expressed functions acquired by a parasitic plant from its host [251].

Infectious bacterial endosymbionts and pathogens are widely considered as potential vectors for horizontal transfer between multicellular organisms [252–256]. Investigators have documented endosymbiont transfers between different multicellular host species [257–259], and many bacteria known as vertebrate pathogens also infect lower eukaryotes, especially amoebae [260–266]. Among these multivalent infectious bacteria, a number are capable of taking up DNA from their immediate environment [267–270].

### 6.5. Viral Integrations into Host Genomes

Viruses of all kinds (including RNA viruses) insert their genomes into eukaryotic host genomes with surprisingly high frequency [271–288]. Integration can occur by retroviral integrase functions, sometimes followed by recombination with other viral sequences [289], or by non-homologous end-joining (NHEJ) at DNA breaks [290,291]. Integration events at DNA breaks have the potential to generate novel sequence configurations.

### 6.6. The Amoeba-Megavirus “Melting Pot” of Sequences from All Three Cell Kingdoms

Viruses have long been considered both as substrates for evolutionary innovation [292,293] and as vectors for horizontal DNA transfer. Particular attention has recently focused on a group of Nucleocytoplasmic Large DNA Viruses (NCLDV) with genomes comprising hundreds of thousands or millions of base-pairs [294–296]. Most significantly, NCLDVs acquire cellular genome fragments and have been found to carry a mixture of DNA sequences from all three major domains of life ([http://shapiro.bsd.uchicago.edu/Viral\\_Composites.html](http://shapiro.bsd.uchicago.edu/Viral_Composites.html)) [297–300]. NCLDVs can infect both protists and multicellular hosts and thus transfer the incorporated cellular sequences. Amoebae are common hosts for many of these large DNA viruses, and amoebae have consequently been designated to constitute an evolutionary “melting pot” ([http://shapiro.bsd.uchicago.edu/Amoebal\\_Viruses.html](http://shapiro.bsd.uchicago.edu/Amoebal_Viruses.html)) [301]. The designation is especially appropriate for two reasons: (1) amoebae are phagocytic and can acquire DNA sequences from engulfed cells [302] and (2) amoebae are hosts to bacteria that both exchange DNA [303] and infect more complex eukaryotes, including both plants and animals [260–266].

Combining the phenomenology of viral infection, amoebal phagocytosis, multivalent bacterial infectivity and endosymbioses with the documentation of viral and prokaryote to eukaryote DNA transfers, it is more than clear that multiple pathways exist by which cells can acquire and transmit DNA segments from one eukaryotic host to another. Specific biochemical activities are distributed among distantly related domains of life [304,305], and we can expect further genomically-documented examples of horizontally acquired adaptations to multiply with continued genome sequencing.

## 7. Protein Evolution by Exon Shuffling and Exonization from “Non-Coding” DNA

Among the most striking results of genomics was the realization that many proteins and the DNA that encodes them are not continuous unitary structures. Many proteins consist of strings of functionally different but interacting “domains,” each one of which may be found iterated in distinct proteins [306–308]. Correspondingly, the cognate protein-coding regions are often discontinuous and composed of expressed coding segments (“exons”) separated by intervening segments (“introns”) [309,310]. While not always the case, DNA exons tended to encode functional protein domains or subdomains [311].

The segmented nature of proteins and protein-coding DNA has major implications for genome functionality and protein evolution:

- A given genetic locus can encode multiple protein products by joining different combinations of exons into the final mRNA by alternative splicing [312–319];
- New protein functionalities can arise rapidly by Lego-like assembly of exons from different sources, known as exon or domain shuffling [320–323];
- Totally new protein domains can originate rapidly by conversion of non-coding DNA segments into exons (“exonization”) and contribute to novel biochemical functions by subsequent duplication and exon shuffling [308,324–326].

Both the rapid evolution of new functionalities by exon shuffling and the origination of sequences encoding extended domains by exonization have potential for protein innovation beyond what is possible through codon-by-codon changes to existing proteins. Exon shuffling has an inherently high

probability of producing adaptive novelties because it rearranges previously evolved sequences that encode established protein functionalities. This potential has been exploited in biotechnology where domain shuffling has proved an efficient method of protein engineering [327–329].

Mobile DNA elements play major roles in both exon shuffling and exonization. The genomic record indicates transposons and retrotransposons can incorporate and relocate exons [330–337], and mobile DNA-mediated exon shuffling has been studied in real time [338–340]. Genomics reveals numerous instances of exonization from mobile element insertions in humans and other mammals [308,326,341–346] as well as in plants [325].

## 8. Regulatory Signal Evolution Involving Mobile DNA Elements

The involvement of mobile DNA in genome change marks one of the most basic divergences between Dobzhansky's Modern Synthesis perspective and a genomics-based view of the evolutionary process. The Modern Synthesis focused on isolated allelic changes at individual loci [347]. Mobile elements, on the other hand, are distributed at many sites throughout the genome and have the potential to generate coordinated changes rewiring distributed regulatory networks involving many loci (<http://shapiro.bsd.uchicago.edu/Table5C-1.MobileElementsFoundtobeExaptedascis-RegulatoryControlSitesinAnimals.html>) [8,348–356].

Genomics documents at least three episodes of regulatory innovation involving mobile elements in the course of vertebrate evolution [357,358], and mobile DNA has been a major source of regulatory motifs in human genome evolution [359,360]. Moreover, the fact that mobile elements are often the most taxonomically specific genome components potentially confers distinctive evolutionary trajectories on different lineages [361–363].

## 9. Adaptations and Innovations in Mammalian Reproduction Arising by Natural Genetic Engineering Processes Involving Mobile DNA and “Non-Coding” ncRNA Molecules

Because of their medical relevance, reproductive biology, embryonic development and stem cell biology in mammals have received particular attention from genomicists. The analysis has uncovered major roles for mobile DNA elements and ncRNAs derived from them. Rather than serving as “fossils that litter our genomes” [364], as conventional evolutionary thinking would assert, these elements are both essential evolutionary tools and active participants in contemporary genome function.

### 9.1. Retroviral Involvement in Placenta Evolution

Mammalian reproduction depends upon development of a syncytial placenta from the zygote to nourish the fetus during pregnancy. At repeated stages in mammalian evolution, distinct endogenous retroviral “envelope” (Env) proteins have been exapted as fusionogenic “syncytins” involved in forming placental tissue in different mammalian lineages ([http://shapiro.bsd.uchicago.edu/Retroviral\\_involvement\\_in\\_placenta\\_evolution.html](http://shapiro.bsd.uchicago.edu/Retroviral_involvement_in_placenta_evolution.html)) [365–370]. Moreover, endogenous retroviruses (ERVs) provide transcriptional regulatory signals to direct imprinted expression of other proteins, such as insulin-like growth factor, required for placental function [371–373].

### 9.2. Mobile DNA Recruitment of Maternal Functions

The endometrium is the maternal tissue that nourishes the placenta in mammalian pregnancy. Endometrial development in the uterus involves the hormone-regulated expression of over 1,500 different proteins. The transcriptional regulatory signals coordinating biogenesis of this pregnancy-specific cohort evolved mainly from mobile DNA elements, both transposons and retrotransposons [374–377]. Convergent patterns of regulatory rewiring can be traced in the endometria of distinct mammalian lineages with well-sequenced genomes [378]. Thus, evolutionary innovations for both fetal and maternal sides of viviparous reproduction arose, to a large degree, through the ability of mammalian cells to mobilize repetitive components of their genomes to adaptive locations.



### 9.3. Mobile DNA and lncRNAs in Stem Cell Programming and Early Embryogenesis

Transcripts from human endogenous retroviruses (HERVs) have been found to be the most stage-specific RNAs expressed during early human embryonic development [379], and there is intrinsic retroviral reactivation in human pre-implantation embryos and pluripotent stem cells [19,380]. Although functional studies are not possible in human embryos, direct functionality has been established for a retroviral RNA in mouse, where MuERV-L transcripts expressed just 8–10 h after fertilization at the 2-cell stage are necessary for developmental competence at the 4-cell stage but not afterwards [381].

HERVs and other mobile DNA elements are the major components of long non-coding lncRNAs, which play important roles in (re)programming embryonic and stem cell genomes (“83% of lncRNAs contain at least one TE (transposable element), while of the total number of base pairs that comprise lncRNA sequences, 42% is derived from TEs” [382]). These include the lncRNA ROR (“regulator of reprogramming”) beginning with a HERV-H transcript needed for formation of human induced pluripotent stem cells (HiPSCs) [383,384], plus LINC01108 (Linc-ES3) and human L1TD1 lncRNAs required to maintain stem cell pluripotency [385,386]. A recent genomic census of over 4000 human-specific binding sites for the transcription factors which reprogram HiPSCs remarkably found between 99.8% and 100% of the sites to be located in mobile DNA repeats [387].

There is a burgeoning literature relating mobile DNA to the evolution of ncRNA-based circuitry essential for mammalian (and, more specifically, human) stem cell and embryonic development [351,375,384,385,388–394]. The lncRNAs bind to and tether a variety of epigenetic modification complexes that execute genome reprogramming [18,395], and the mobile DNA element sequences in each molecule have been proposed to constitute a combinatorial code of RNA domains that link together different genome modification processes (Called the “RIDL hypothesis” for Repeat Insertion Domains of lncRNAs) [394]. In addition to lncRNAs, mobile DNA elements have also been documented to be sources for cell regulatory miRNAs [396–399].

## 10. A 21st Century Evolutionary Principle: Cell-Mediated Variation of Read-Write (RW) Genomes

To recapitulate, a genomics-based view of evolutionary variation introduces novel features to hereditary control of cell biology impossible to predict when Dobzhansky and other evolutionary biologists formulated the neo-Darwinian Modern Synthesis in the middle of the last century:

- The existence of three distinct realms of cell evolution, Bacteria, Archaea and Eukarya;
- Symbiogenetic fusions involving these different realms leading to the formation of eukaryotic cells bearing organelles with multiple genome compartments;
- Horizontal organelle, virus and DNA transfers affecting adaptive traits across all cell types;
- The functional organization of proteins as systems of distinct interacting domains encoded by exons and subject to rapid evolution by exon shuffling and exon origination from non-coding DNA (exonization);
- Establishment of adaptive, distributed genome networks integrated by mobile DNA elements dispersing repetitive regulatory signals to multiple loci;
- Regulation of cell differentiation in multicellular development by non-coding lncRNA molecules composed largely of mobile repetitive DNA elements that serve as scaffolds for epigenetic modifying activities.

Altogether, the combinatorial coding and regulatory aspects of cell heredity, plus the biochemical abilities cells possess to rearrange DNA molecules, constitute a powerful toolbox for adaptive genome rewriting. Revelations from genomic analysis oblige us to reconsider the simplifying assumptions made in the past two centuries about the nature of evolutionary variation. Rather than single gene traits, we recognize that all phenotypes involve coordinated activity by multiple interacting cell molecules.

As summarized above, we have evidence that genomes contain abundant and functional repetitive components in addition to the unique coding sequences envisaged in the early days of molecular biology [12]. Instead of the “Constant Genome,” subject to accidental modification, we know today that cells possess “Read–Write Genomes” they can alter by numerous biochemical processes capable of rapidly restructuring cellular DNA molecules [6,8]. Genomics has modernized our understanding of the evolutionary process. Rather than viewing genome evolution as a happenstance series of copying errors, we are now in a position to study it as a complex biological process of active self-modification.

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## References

1. Ayala, F.J. Nothing in biology makes sense except in the light of evolution: Theodosius Dobzhansky: 1900–1975. *J. Hered.* **1977**, *68*, 3–10. [[PubMed](#)]
2. Dobzhansky, T. Nothing in biology makes sense except in the light of evolution. *Am. Biol. Teach.* **1973**, *35*, 125–129. [[CrossRef](#)]
3. Watson, J.D.; Crick, F.H. Genetical implications of the structure of deoxyribonucleic acid. *Nature* **1953**, *171*, 964–967. [[CrossRef](#)] [[PubMed](#)]
4. Alberts, B.; Johnson, A.; Lewis, J.; Raff, M.; Roberts, K.; Walter, P. *Molecular Biology of the Cell*; Garland Science: New York, NY, USA, 2002.
5. Darwin, C. *Origin of Species*; John Russel: London, UK, 1859.
6. Shapiro, J.A. *Evolution: A View from the 21st Century*; FT Press Science: Upper Saddle River, NJ, USA, 2011.
7. Shapiro, J.A. Constraint and opportunity in genome innovation. *RNA Biol.* **2014**, *11*, 186–196. [[CrossRef](#)] [[PubMed](#)]
8. Shapiro, J.A. How life changes itself: The Read-Write (RW) genome. *Phys. Life Rev.* **2013**, *10*, 287–323. [[CrossRef](#)] [[PubMed](#)]
9. Gould, S.J. Punctuated equilibrium and the fossil record. *Science* **1983**, *219*, 439–440. [[CrossRef](#)] [[PubMed](#)]
10. Shapiro, J.A. The basic concept of the read-write genome: Mini-review on cell-mediated DNA modification. *Biosystems* **2016**, *140*, 35–37. [[CrossRef](#)] [[PubMed](#)]
11. Myers, R.M.; Stamatoyannopoulos, J.; Snyder, M.; Dunham, I.; Hardison, R.C.; Bernstein, B.E.; Gingeras, T.R.; Kent, W.J.; Birney, E.; Wold, B.; *et al.* A user’s guide to the encyclopedia of DNA elements (ENCODE). *PLoS Biol.* **2011**, *9*, e1001046.
12. Shapiro, J.A.; Sternberg, R.V. Why repetitive DNA is essential to genome function. *Biol. Rev. Camb. Philos. Soc.* **2005**, *80*, 227–250. [[CrossRef](#)]
13. De Koning, A.P.; Gu, W.; Castoe, T.A.; Batzer, M.A.; Pollock, D.D. Repetitive elements may comprise over two-thirds of the human genome. *PLoS Genet.* **2011**, *7*, e1002384. [[CrossRef](#)] [[PubMed](#)]
14. Wang, S.; Tran, E.J. Unexpected functions of lncRNAs in gene regulation. *Commun. Integr. Biol.* **2013**, *6*, e27610. [[CrossRef](#)] [[PubMed](#)]
15. Huarte, M. lncRNAs have a say in protein translation. *Cell Res.* **2013**, *23*, 449–451. [[CrossRef](#)] [[PubMed](#)]
16. Necsulea, A.; Soumillon, M.; Warnefors, M.; Liechti, A.; Daish, T.; Zeller, U.; Baker, J.C.; Grützner, F.; Kaessmann, H. The evolution of lncRNA repertoires and expression patterns in tetrapods. *Nature* **2014**, *505*, 635–640. [[CrossRef](#)] [[PubMed](#)]
17. Guan, D.; Zhang, W.; Zhang, W.; Liu, G.H.; Belmonte, J.C. Switching cell fate, ncRNAs coming to play. *Cell Death Dis.* **2013**, *4*. [[CrossRef](#)] [[PubMed](#)]
18. Guttman, M.; Donaghey, J.; Carey, B.W.; Garber, M.; Grenier, J.K.; Munson, G.; Young, G.; Lucas, A.B.; Ach, R.; Bruhn, L.; *et al.* lincRNAs act in the circuitry controlling pluripotency and differentiation. *Nature* **2011**, *477*, 295–300. [[CrossRef](#)] [[PubMed](#)]
19. St Laurent, G.; Shtokalo, D.; Dong, B.; Tackett, M.R.; Fan, X.; Lazorthes, S.; Nicolas, E.; Sang, N.; Triche, T.J.; McCaffrey, T.A.; *et al.* VlinRNAs controlled by retroviral elements are a hallmark of pluripotency and cancer. *Genome Biol.* **2013**, *14*. [[CrossRef](#)] [[PubMed](#)]

20. Carroll, S.B. Evo-devo and an expanding evolutionary synthesis: A genetic theory of morphological evolution. *Cell* **2008**, *134*, 25–36. [[CrossRef](#)] [[PubMed](#)]
21. Prud'homme, B.; Gompel, N.; Carroll, S.B. Emerging principles of regulatory evolution. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 8605–8612. [[CrossRef](#)] [[PubMed](#)]
22. Sogin, S.J.; Sogin, M.L.; Woese, C.R. Phylogenetic measurement in prokaryotes by primary structural characterization. *J. Mol. Evol.* **1971**, *1*, 173–184. [[CrossRef](#)] [[PubMed](#)]
23. Woese, C.R.; Fox, G.E. Phylogenetic structure of the prokaryotic domain: The primary kingdoms. *Proc. Natl. Acad. Sci. USA* **1977**, *74*, 5088–5090. [[CrossRef](#)] [[PubMed](#)]
24. Woese, C.R.; Magrum, L.J.; Fox, G.E. Archaeobacteria. *J. Mol. Evol.* **1978**, *11*, 245–251. [[CrossRef](#)] [[PubMed](#)]
25. Kyrpides, N.C.; Woese, C.R. Universally conserved translation initiation factors. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 224–228. [[CrossRef](#)] [[PubMed](#)]
26. Huxley, J. *Evolution: The Modern Synthesis*; Allen & Unwin: London, UK, 1942.
27. Mayr, E. *The Growth of Biological Thought: Diversity, Evolution, and Inheritance*; Belknap Press: Cambridge, MA, USA, 1982.
28. Albers, S.V.; Forterre, P.; Prangishvili, D.; Schleper, C. The legacy of Carl Woese and Wolfram Zillig: From phylogeny to landmark discoveries. *Nat. Rev. Microbiol.* **2013**, *11*, 713–719. [[CrossRef](#)] [[PubMed](#)]
29. Sapp, J. *The New Foundations of Evolution: On the Tree of Life*; Oxford University Press: Oxford, UK, 2009.
30. Woese, C.R. On the evolution of cells. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 8742–8747. [[CrossRef](#)] [[PubMed](#)]
31. Woese, C.R. Archaeobacteria. *Sci. Am.* **1981**, *244*, 98–122. [[CrossRef](#)]
32. Margulis, L. *Symbiosis in Cell Evolution*; W.H. Freeman Co.: London, UK, 1981.
33. Margulis, L. Symbiosis and evolution. *Sci. Am.* **1971**, *225*, 48–57. [[CrossRef](#)] [[PubMed](#)]
34. Margulis, L.; Sagan, D. *Acquiring Genomes: A Theory of the Origins of Species*; Perseus Books Group: Amherst, MA, USA, 2002.
35. Margulis, L. *Origin of Eukaryotic Cells*; Yale University Press: New Haven, CT, USA, 1970.
36. Foster, P.G.; Cox, C.J.; Embley, T.M. The primary divisions of life: A phylogenomic approach employing composition-heterogeneous methods. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2009**, *364*, 2197–2207. [[CrossRef](#)] [[PubMed](#)]
37. Embley, T.M.; Martin, W. Eukaryotic evolution, changes and challenges. *Nature* **2006**, *440*, 623–630. [[CrossRef](#)] [[PubMed](#)]
38. Lithgow, T.; Schneider, A. Evolution of macromolecular import pathways in mitochondria, hydrogenosomes and mitosomes. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2010**, *365*, 799–817. [[CrossRef](#)] [[PubMed](#)]
39. Shiflett, A.M.; Johnson, P.J. Mitochondrion-related organelles in eukaryotic protists. *Ann. Rev. Microbiol.* **2010**, *64*, 409–429. [[CrossRef](#)] [[PubMed](#)]
40. Woese, C.R. Endosymbionts and mitochondrial origins. *J. Mol. Evol.* **1977**, *10*, 93–96. [[CrossRef](#)] [[PubMed](#)]
41. Esser, C.; Ahmadinejad, N.; Wiegand, C.; Rotte, C.; Sebastiani, F.; Gelius-Dietrich, G.; Henze, K.; Kretschmann, E.; Richly, E.; Leister, D.; *et al.* A genome phylogeny for mitochondria among alpha-proteobacteria and a predominantly eubacterial ancestry of yeast nuclear genes. *Mol. Biol. Evol.* **2004**, *21*, 1643–1660. [[CrossRef](#)] [[PubMed](#)]
42. Vesteg, M.; Krajcovic, J. Origin of eukaryotic cells as a symbiosis of parasitic alpha-proteobacteria in the periplasm of two-membrane-bounded sexual pre-karyotes. *Commun. Integr. Biol.* **2008**, *1*, 104–113. [[CrossRef](#)] [[PubMed](#)]
43. Zablen, L.B.; Kissil, M.S.; Woese, C.R.; Buetow, D.E. Phylogenetic origin of the chloroplast and prokaryotic nature of its ribosomal RNA. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 2418–2422. [[CrossRef](#)] [[PubMed](#)]
44. Bonen, L.; Doolittle, W.F. On the prokaryotic nature of red algal chloroplasts. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 2310–2314. [[CrossRef](#)] [[PubMed](#)]
45. Green, B.R. Chloroplast genomes of photosynthetic eukaryotes. *Plant J.* **2011**, *66*, 34–44. [[CrossRef](#)] [[PubMed](#)]
46. Cavalier-Smith, T. Chloroplast evolution: Secondary symbiogenesis and multiple losses. *Curr. Biol.* **2002**, *12*, R62–R64. [[CrossRef](#)]
47. Krause, K. From chloroplasts to “cryptic” plastids: Evolution of plastid genomes in parasitic plants. *Curr. Genet.* **2008**, *54*, 111–121. [[CrossRef](#)] [[PubMed](#)]
48. Thorsness, P.E.; Fox, T.D. Escape of DNA from mitochondria to the nucleus in *Saccharomyces cerevisiae*. *Nature* **1990**, *346*, 376–379. [[CrossRef](#)] [[PubMed](#)]

49. Hazkani-Covo, E. Mitochondrial insertions into primate nuclear genomes suggest the use of numts as a tool for phylogeny. *Mol. Biol. Evol.* **2009**, *26*, 2175–2179. [[CrossRef](#)] [[PubMed](#)]
50. Bachtrog, D.; Hornton, K.; Clark, A.; Andolfatto, P. Extensive introgression of mitochondrial DNA relative to nuclear genes in the *Drosophila yakuba* species group. *Evolution* **2006**, *60*, 292–302. [[CrossRef](#)] [[PubMed](#)]
51. Adams, K.L.; Qiu, Y.L.; Stoutemyer, M.; Palmer, J.D. Punctuated evolution of mitochondrial gene content: High and variable rates of mitochondrial gene loss and transfer to the nucleus during angiosperm evolution. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 9905–9912. [[CrossRef](#)] [[PubMed](#)]
52. Stegemann, S.; Hartmann, S.; Ruf, S.; Bock, R. High-frequency gene transfer from the chloroplast genome to the nucleus. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 8828–8833. [[CrossRef](#)] [[PubMed](#)]
53. Rousseau-Gueutin, M.; Ayliffe, M.A.; Timmis, J.N. Conservation of plastid sequences in the plant nuclear genome for millions of years facilitates endosymbiotic evolution. *Plant Physiol.* **2011**, *157*, 2181–2193. [[CrossRef](#)] [[PubMed](#)]
54. Keeling, P.J. The endosymbiotic origin, diversification and fate of plastids. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2010**, *365*, 729–748. [[CrossRef](#)] [[PubMed](#)]
55. Bock, R.; Timmis, J.N. Reconstructing evolution: Gene transfer from plastids to the nucleus. *Bioessays* **2008**, *30*, 556–566. [[CrossRef](#)] [[PubMed](#)]
56. Ricchetti, M.; Tekaiia, F.; Dujon, B. Continued colonization of the human genome by mitochondrial DNA. *PLoS Biol.* **2004**, *2*, e273. [[CrossRef](#)] [[PubMed](#)]
57. Lloyd, A.H.; Timmis, J.N. Endosymbiotic evolution in action: Real-time observations of chloroplast to nucleus gene transfer. *Mob. Genet. Elem.* **2011**, *1*, 216–220. [[CrossRef](#)] [[PubMed](#)]
58. Huang, C.Y.; Ayliffe, M.A.; Timmis, J.N. Direct measurement of the transfer rate of chloroplast DNA into the nucleus. *Nature* **2003**, *422*, 72–76. [[CrossRef](#)] [[PubMed](#)]
59. Cullis, C.A.; Vorster, B.J.; van der Vyver, C.; Kunert, K.J. Transfer of genetic material between the chloroplast and nucleus: How is it related to stress in plants? *Ann. Bot.* **2009**, *103*, 625–633. [[CrossRef](#)] [[PubMed](#)]
60. Roark, L.M.; Hui, A.Y.; Donnelly, L.; Birchler, J.A.; Newton, K.J. Recent and frequent insertions of chloroplast DNA into maize nuclear chromosomes. *Cytogenet. Genome Res.* **2010**, *129*, 17–23. [[CrossRef](#)] [[PubMed](#)]
61. Cheng, X.; Ivessa, A.S. The migration of mitochondrial DNA fragments to the nucleus affects the chronological aging process of *Saccharomyces cerevisiae*. *Aging Cell* **2010**, *9*, 919–923. [[CrossRef](#)] [[PubMed](#)]
62. Caro, P.; Gómez, J.; Arduini, A.; González-Sánchez, M.; González-García, M.; Borrás, C.; Viña, J.; Puertas, M.J.; Sastre, J.; Barja, G. Mitochondrial DNA sequences are present inside nuclear DNA in rat tissues and increase with age. *Mitochondrion* **2010**, *10*, 479–486. [[CrossRef](#)] [[PubMed](#)]
63. Hazkani-Covo, E.; Zeller, R.M.; Martin, W. Molecular poltergeists: Mitochondrial DNA copies (*numts*) in sequenced nuclear genomes. *PLoS Genet.* **2010**, *6*, e1000834. [[CrossRef](#)] [[PubMed](#)]
64. Ju, Y.S.; Tubio, J.M.; Mifsud, W.; Fu, B.; Davies, H.R.; Ramakrishna, M.; Li, Y.; Yates, L.; Gundem, G.; Tarpey, P.S.; *et al.* Frequent somatic transfer of mitochondrial DNA into the nuclear genome of human cancer cells. *Genome Res.* **2015**, *25*, 814–824. [[CrossRef](#)] [[PubMed](#)]
65. Ricchetti, M.; Fairhead, C.; Dujon, B. Mitochondrial DNA repairs double-strand breaks in yeast chromosomes. *Nature* **1999**, *402*, 96–100. [[CrossRef](#)] [[PubMed](#)]
66. Rodriguez-Moreno, L.; González, V.M.; Benjak, A.; Martí, M.C.; Puigdomènech, P.; Aranda, M.A.; Garcia-Mas, J. Determination of the melon chloroplast and mitochondrial genome sequences reveals that the largest reported mitochondrial genome in plants contains a significant amount of DNA having a nuclear origin. *BMC Genom.* **2011**, *12*. [[CrossRef](#)] [[PubMed](#)]
67. Petrokovski, S.; Trifonov, E.N. Imported sequences in the mitochondrial yeast genome identified by nucleotide linguistics. *Gene* **1992**, *122*, 129–137. [[CrossRef](#)]
68. Adams, K.L.; Daley, D.O.; Whelan, J.; Palmer, J.D. Genes for two mitochondrial ribosomal proteins in flowering plants are derived from their chloroplast or cytosolic counterparts. *Plant Cell* **2002**, *14*, 931–943. [[CrossRef](#)] [[PubMed](#)]
69. Koulintchenko, M.; Konstantinov, Y.; Dietrich, A. Plant mitochondria actively import DNA via the permeability transition pore complex. *EMBO J.* **2003**, *22*, 1245–1254. [[CrossRef](#)] [[PubMed](#)]
70. Nozaki, H.; Matsuzaki, M.; Takahara, M.; Misumi, O.; Kuroiwa, H.; Hasegawa, M.; Shin-i, T.; Kohara, Y.; Ogasawara, N.; Kuroiwa, T. The phylogenetic position of red algae revealed by multiple nuclear genes from mitochondria-containing eukaryotes and an alternative hypothesis on the origin of plastids. *J. Mol. Evol.* **2003**, *56*, 485–497. [[CrossRef](#)] [[PubMed](#)]

71. Jeyaprakash, A.; Hoy, M.A. First divergence time estimate of spiders, scorpions, mites and ticks (subphylum: Chelicerata) inferred from mitochondrial phylogeny. *Exp. Appl. Acarol.* **2009**, *47*, 1–18. [[CrossRef](#)] [[PubMed](#)]
72. Bullerwell, C.E.; Gray, M.W. Evolution of the mitochondrial genome: Protist connections to animals, fungi and plants. *Curr. Opin. Microbiol.* **2004**, *7*, 528–534. [[CrossRef](#)] [[PubMed](#)]
73. Burger, G.; Gray, M.W.; Lang, B.F. Mitochondrial genomes: Anything goes. *Trends Genet.* **2003**, *19*, 709–716. [[CrossRef](#)] [[PubMed](#)]
74. Gray, M.W.; Lang, B.F.; Burger, G. Mitochondria of protists. *Ann. Rev. Genet.* **2004**, *38*, 477–524. [[CrossRef](#)] [[PubMed](#)]
75. Gray, M.W.; Burger, G.; Lang, B.F. The origin and early evolution of mitochondria. *Genome Biol.* **2001**, *2*. [[CrossRef](#)]
76. Gray, M.W.; Burger, G.; Lang, B.F. Mitochondrial evolution. *Science* **1999**, *283*, 1476–1481. [[CrossRef](#)] [[PubMed](#)]
77. Sanchez-Puerta, M.V.; Cho, Y.; Mower, J.P.; Alverson, A.J.; Palmer, J.D. Frequent, phylogenetically local horizontal transfer of the *cox1* group I Intron in flowering plant mitochondria. *Mol. Biol. Evol.* **2008**, *25*, 1762–1777. [[CrossRef](#)] [[PubMed](#)]
78. Hikosaka, K.; Watanabe, Y.; Tsuji, N.; Kita, K.; Kishine, H.; Arisue, N.; Palacpac, N.M.; Kawazu, S.; Sawai, H.; Horii, T.; *et al.* Divergence of the mitochondrial genome structure in the apicomplexan parasites, *Babesia* and *Theileria*. *Mol. Biol. Evol.* **2010**, *27*, 1107–1116. [[CrossRef](#)] [[PubMed](#)]
79. Hikosaka, K.; Watanabe, Y.; Kobayashi, F.; Waki, S.; Kita, K.; Tanabe, K. Highly conserved gene arrangement of the mitochondrial genomes of 23 *Plasmodium* species. *Parasitol. Int.* **2011**, *60*, 175–180. [[CrossRef](#)] [[PubMed](#)]
80. Valach, M.; Farkas, Z.; Fricova, D.; Kovac, J.; Brejova, B.; Vinar, T.; Pfeiffer, I.; Kucsera, J.; Tomaska, L.; Lang, B.F.; *et al.* Evolution of linear chromosomes and multipartite genomes in yeast mitochondria. *Nucleic Acids Res.* **2011**, *39*, 4202–4219. [[CrossRef](#)] [[PubMed](#)]
81. Smith, D.R.; Kayal, E.; Yanagihara, A.A.; Collins, A.G.; Pirro, S.; Keeling, P.J. First complete mitochondrial genome sequence from a box jellyfish reveals a highly fragmented linear architecture and insights into telomere evolution. *Genome Biol. Evol.* **2012**, *4*, 52–58. [[CrossRef](#)] [[PubMed](#)]
82. Kayal, E.; Bentlage, B.; Collins, A.G.; Kayal, M.; Pirro, S.; Lavrov, D.V. Evolution of linear mitochondrial genomes in medusozoan cnidarians. *Genome Biol. Evol.* **2012**, *4*, 1–12. [[CrossRef](#)] [[PubMed](#)]
83. Sloan, D.B.; Alverson, A.J.; Chuckalovcak, J.P.; Wu, M.; McCauley, D.E.; Palmer, J.D.; Taylor, D.R. Rapid evolution of enormous, multichromosomal genomes in flowering plant mitochondria with exceptionally high mutation rates. *PLoS Biol.* **2012**, *10*, e1001241. [[CrossRef](#)] [[PubMed](#)]
84. Sloan, D.B.; Alverson, A.J.; Wu, M.; Palmer, J.D.; Taylor, D.R. Recent acceleration of plastid sequence and structural evolution coincides with extreme mitochondrial divergence in the angiosperm genus *Silene*. *Genome Biol. Evol.* **2012**, *4*, 294–306. [[CrossRef](#)] [[PubMed](#)]
85. Hjort, K.; Goldberg, A.V.; Tsaousis, A.D.; Hirt, R.P.; Embley, T.M. Diversity and reductive evolution of mitochondria among microbial eukaryotes. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2010**, *365*, 713–727. [[CrossRef](#)] [[PubMed](#)]
86. Marande, W.; Lukes, J.; Burger, G. Unique mitochondrial genome structure in diplomonads, the sister group of kinetoplastids. *Eukaryot. Cell* **2005**, *4*, 1137–1146. [[CrossRef](#)] [[PubMed](#)]
87. Brown, W.M.; George, M., Jr.; Wilson, A.C. Rapid evolution of animal mitochondrial DNA. *Proc. Natl. Acad. Sci. USA* **1979**, *76*, 1967–1971. [[CrossRef](#)] [[PubMed](#)]
88. Embley, T.M. Multiple secondary origins of the anaerobic lifestyle in eukaryotes. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2006**, *361*, 1055–1067. [[CrossRef](#)] [[PubMed](#)]
89. Van der Giezen, M. Hydrogenosomes and mitosomes: Conservation and evolution of functions. *J. Eukaryot. Microbiol.* **2009**, *56*, 221–231. [[CrossRef](#)] [[PubMed](#)]
90. Hackstein, J.H.; Tjaden, J.; Huynen, M. Mitochondria, hydrogenosomes and mitosomes: Products of evolutionary tinkering! *Curr. Genet.* **2006**, *50*, 225–245. [[CrossRef](#)] [[PubMed](#)]
91. Brouard, J.S.; Otis, C.; Lemieux, C.; Turmel, M. The exceptionally large chloroplast genome of the green alga *Floydiella terrestris* illuminates the evolutionary history of the *Chlorophyceae*. *Genome Biol. Evol.* **2010**, *2*, 240–256. [[CrossRef](#)] [[PubMed](#)]
92. Magee, A.M.; Aspinall, S.; Rice, D.W.; Cusack, B.P.; Sémon, M.; Perry, A.S.; Stefanović, S.; Milbourne, D.; Barth, S.; Palmer, J.D.; *et al.* Localized hypermutation and associated gene losses in legume chloroplast genomes. *Genome Res.* **2010**, *20*, 1700–1710. [[CrossRef](#)] [[PubMed](#)]



93. Wu, C.S.; Lin, C.P.; Hsu, C.Y.; Wang, R.J.; Chaw, S.M. Comparative chloroplast genomes of pinaceae: Insights into the mechanism of diversified genomic organizations. *Genome Biol. Evol.* **2011**, *3*, 309–319. [[CrossRef](#)] [[PubMed](#)]
94. Wolf, P.G.; Der, J.P.; Duffy, A.M.; Davidson, J.B.; Grusz, A.L.; Pryer, K.M. The evolution of chloroplast genes and genomes in ferns. *Plant Mol. Biol.* **2011**, *76*, 251–261. [[CrossRef](#)] [[PubMed](#)]
95. Reyes-Prieto, A.; Yoon, H.S.; Moustafa, A.; Yang, E.C.; Andersen, R.A.; Boo, S.M.; Nakayama, T.; Ishida, K.I.; Bhattacharya, D. Differential gene retention in plastids of common recent origin. *Mol. Biol. Evol.* **2010**, *27*, 1530–1537. [[CrossRef](#)] [[PubMed](#)]
96. Gould, S.B.; Waller, R.F.; McFadden, G.I. Plastid evolution. *Ann. Rev. Plant Biol.* **2008**, *59*, 491–517. [[CrossRef](#)] [[PubMed](#)]
97. Smith, D.R.; Lee, R.W. A plastid without a genome: Evidence from the nonphotosynthetic green alga *Polytomella*. *Plant Physiol.* **2014**, *164*, 1812–1819. [[CrossRef](#)] [[PubMed](#)]
98. Stiller, J.W.; Huang, J.; Ding, Q.; Tian, J.; Goodwillie, C. Are algal genes in nonphotosynthetic protists evidence of historical plastid endosymbioses? *BMC Genom.* **2009**, *10*. [[CrossRef](#)] [[PubMed](#)]
99. Revill, M.J.; Stanley, S.; Hibberd, J.M. Plastid genome structure and loss of photosynthetic ability in the parasitic genus *Cuscuta*. *J. Exp. Bot.* **2005**, *56*, 2477–2486. [[CrossRef](#)] [[PubMed](#)]
100. Barbrook, A.C.; Howe, C.J.; Purton, S. Why are plastid genomes retained in non-photosynthetic organisms? *Trends Plant Sci.* **2006**, *11*, 101–108. [[CrossRef](#)] [[PubMed](#)]
101. Baurain, D.; Brinkmann, H.; Petersen, J.; Rodríguez-Ezpeleta, N.; Stechmann, A.; Demoulin, V.; Roger, A.J.; Burger, G.; Lang, B.F.; Philippe, H. Phylogenomic evidence for separate acquisition of plastids in cryptophytes, haptophytes, and stramenopiles. *Mol. Biol. Evol.* **2010**, *27*, 1698–1709. [[CrossRef](#)] [[PubMed](#)]
102. Keeling, P.J. Chromalveolates and the evolution of plastids by secondary endosymbiosis. *J. Eukaryot. Microbiol.* **2009**, *56*, 1–8. [[CrossRef](#)] [[PubMed](#)]
103. Archibald, J.M. Plastid evolution: Remnant algal genes in ciliates. *Curr. Biol.* **2008**, *18*, R663–R665. [[CrossRef](#)] [[PubMed](#)]
104. Kutschera, U.; Niklas, K.J. Macroevolution via secondary endosymbiosis: A Neo-Goldschmidtian view of unicellular hopeful monsters and Darwin's primordial intermediate form. *Theory Biosci.* **2008**, *127*, 277–289. [[CrossRef](#)] [[PubMed](#)]
105. Zauner, S.; Lockhart, P.; Stoebe-Maier, B.; Gilson, P.; McFadden, G.I.; Maier, U.G. Differential gene transfers and gene duplications in primary and secondary endosymbioses. *BMC Evol. Biol.* **2006**, *6*. [[CrossRef](#)] [[PubMed](#)]
106. Janouškovec, J.; Horák, A.; Oborník, M.; Lukeš, J.; Keeling, P.J. A common red algal origin of the apicomplexan, dinoflagellate, and heterokont plastids. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 10949–10954. [[CrossRef](#)] [[PubMed](#)]
107. Moore, C.E.; Archibald, J.M. Nucleomorph genomes. *Ann. Rev. Genet.* **2009**, *43*, 251–264. [[CrossRef](#)] [[PubMed](#)]
108. Silver, T.D.; Koike, S.; Yabuki, A.; Kofuji, R.; Archibald, J.M.; Ishida, K.I. Phylogeny and nucleomorph karyotype diversity of chlorarachniophyte algae. *J. Eukaryot. Microbiol.* **2007**, *54*, 403–410. [[CrossRef](#)] [[PubMed](#)]
109. Archibald, J.M. Nucleomorph genomes: Structure, function, origin and evolution. *Bioessays* **2007**, *29*, 392–402. [[CrossRef](#)] [[PubMed](#)]
110. Maruyama, S.; Sugahara, J.; Kanai, A.; Nozaki, H. Permuted tRNA genes in the nuclear and nucleomorph genomes of photosynthetic eukaryotes. *Mol. Biol. Evol.* **2010**, *27*, 1070–1076. [[CrossRef](#)] [[PubMed](#)]
111. Curtis, B.A.; Tanifuji, G.; Burki, F.; Gruber, A.; Irimia, M.; Maruyama, S.; Arias, M.C.; Ball, S.G.; Gile, G.H.; Hiraoka, Y.; *et al.* Algal genomes reveal evolutionary mosaicism and the fate of nucleomorphs. *Nature* **2012**, *492*, 59–65. [[CrossRef](#)] [[PubMed](#)]
112. Moore, C.E.; Curtis, B.; Mills, T.; Tanifuji, G.; Archibald, J.M. Nucleomorph genome sequence of the cryptophyte alga *Chroomonas mesostigmatica* CCMP1168 reveals lineage-specific gene loss and genome complexity. *Genome Biol. Evol.* **2012**, *4*, 1162–1175. [[CrossRef](#)] [[PubMed](#)]
113. Cavalier-Smith, T. Nucleomorphs: Enslaved algal nuclei. *Curr. Opin. Microbiol.* **2002**, *5*, 612–619. [[CrossRef](#)]
114. Trench, R.K.; Greene, R.W.; Bystrom, B.G. Chloroplasts as functional organelles in animal tissues. *J. Cell Biol.* **1969**, *42*, 404–417. [[CrossRef](#)] [[PubMed](#)]

115. Händeler, K.; Grzybowski, Y.P.; Krug, P.J.; Wägele, H. Functional chloroplasts in metazoan cells—A unique evolutionary strategy in animal life. *Front. Zool.* **2009**, *6*. [[CrossRef](#)] [[PubMed](#)]
116. Serôdio, J.; Cruz, S.; Cartaxana, P.; Calado, R. Photophysiology of kleptoplasts: Photosynthetic use of light by chloroplasts living in animal cells. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2014**, *369*. [[CrossRef](#)] [[PubMed](#)]
117. Pierce, S.K.; Curtis, N.E. Cell biology of the chloroplast symbiosis in sacoglossan sea slugs. *Int. Rev. Cell Mol. Biol.* **2012**, *293*, 123–148. [[PubMed](#)]
118. Baumgartner, F.A.; Pavia, H.; Toth, G.B. Acquired phototrophy through retention of functional chloroplasts increases growth efficiency of the sea slug *Elysia viridis*. *PLoS ONE* **2015**, *10*, e0120874. [[CrossRef](#)] [[PubMed](#)]
119. Wägele, H.; Deusch, O.; Händeler, K.; Martin, R.; Schmitt, V.; Christa, G.; Pinzger, B.; Gould, S.B.; Dagan, T.; Klussmann-Kolb, A.; et al. Transcriptomic evidence that longevity of acquired plastids in the photosynthetic slugs *Elysia timida* and *Plakobranthus ocellatus* does not entail lateral transfer of algal nuclear genes. *Mol. Biol. Evol.* **2011**, *28*, 699–706. [[CrossRef](#)] [[PubMed](#)]
120. Muscatine, L.; Greene, R.W. Chloroplasts and algae as symbionts in molluscs. *Int. Rev. Cytol.* **1973**, *36*, 137–169. [[PubMed](#)]
121. Green, B.J.; Li, W.Y.; Manhart, J.R.; Fox, T.C.; Summer, E.J.; Kennedy, R.A.; Pierce, S.K.; Rumpho, M.E. Mollusc-algal chloroplast endosymbiosis. Photosynthesis, thylakoid protein maintenance, and chloroplast gene expression continue for many months in the absence of the algal nucleus. *Plant Physiol.* **2000**, *124*, 331–342. [[CrossRef](#)] [[PubMed](#)]
122. Gomez, F.; Lopez-Garcia, P.; Moreira, D. Molecular phylogeny of the ocelloid-bearing dinoflagellates *Erythroapsidinium* and *Warnowia* (warnowiaceae, dinophyceae). *J. Eukaryot. Microbiol.* **2009**, *56*, 440–445. [[CrossRef](#)] [[PubMed](#)]
123. Hoppenrath, M.; Bachvaroff, T.R.; Handy, S.M.; Delwiche, C.F.; Leander, B.S. Molecular phylogeny of ocelloid-bearing dinoflagellates (Warnowiaceae) as inferred from SSU and LSU rDNA sequences. *BMC Evol. Biol.* **2009**, *9*. [[CrossRef](#)] [[PubMed](#)]
124. Gavelis, G.S.; Hayakawa, S.; White, R.A., III; Gojobori, T.; Suttle, C.A.; Keeling, P.J.; Leander, B.S. Eye-like ocelloids are built from different endosymbiotically acquired components. *Nature* **2015**, *523*, 204–207. [[CrossRef](#)] [[PubMed](#)]
125. Hulse, J.H.; Spurgeon, D. Triticale. *Sci. Am.* **1974**, *231*, 72–80. [[CrossRef](#)]
126. Bento, M.; Gustafson, P.; Viegas, W.; Silva, M. Genome merger: From sequence rearrangements in triticale to their elimination in wheat-rye addition lines. *Theor. Appl. Genet.* **2010**, *121*, 489–497. [[CrossRef](#)] [[PubMed](#)]
127. Bento, M.; Pereira, H.S.; Rocheta, M.; Gustafson, P.; Viegas, W.; Silva, M. Polyploidization as a retraction force in plant genome evolution: Sequence rearrangements in triticale. *PLoS ONE* **2008**, *3*, e1402. [[CrossRef](#)] [[PubMed](#)]
128. Bento, M.; Gustafson, J.P.; Viegas, W.; Silva, M. Size matters in Triticeae polyploids: Larger genomes have higher remodeling. *Genome* **2011**, *54*, 175–183. [[PubMed](#)]
129. Anderson, E.; Stebbins, G.L., Jr. Hybridization as an evolutionary stimulus. *Evolution* **1954**, *8*, 378–388. [[CrossRef](#)]
130. Stebbins, G.L. Cataclysmic Evolution. *Sci. Am.* **1951**, *184*, 54–59. [[CrossRef](#)]
131. Ungerer, M.C.; Baird, S.J.; Pan, J.; Rieseberg, L.H. Rapid hybrid speciation in wild sunflowers. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 11757–11762. [[CrossRef](#)] [[PubMed](#)]
132. Fuentes, I.; Stegemann, S.; Golczyk, H.; Karcher, D.; Bock, R. Horizontal genome transfer as an asexual path to the formation of new species. *Nature* **2014**, *511*, 232–235. [[CrossRef](#)] [[PubMed](#)]
133. Véron, A.S.; Lemaitre, C.; Gautier, C.; Lacroix, V.; Sagot, M.F. Close 3D proximity of evolutionary breakpoints argues for the notion of spatial synteny. *BMC Genom.* **2011**, *12*. [[CrossRef](#)] [[PubMed](#)]
134. Catchen, J.M.; Conery, J.S.; Postlethwait, J.H. Automated identification of conserved synteny after whole-genome duplication. *Genome Res.* **2009**, *19*, 1497–1505. [[CrossRef](#)] [[PubMed](#)]
135. Tang, H. Synteny and collinearity in plant genomes. *Science* **2008**, *320*, 486–488. [[CrossRef](#)] [[PubMed](#)]
136. Wolfe, K.H.; Shields, D.C. Molecular evidence for an ancient duplication of the entire yeast genome. *Nature* **1997**, *387*, 708–713. [[CrossRef](#)] [[PubMed](#)]
137. Wolfe, K.H. Origin of the yeast whole-genome duplication. *PLoS Biol.* **2015**, *13*, e1002221. [[CrossRef](#)] [[PubMed](#)]

138. Marcet-Houben, M.; Gabaldon, T. Beyond the whole-genome duplication: Phylogenetic evidence for an ancient interspecies hybridization in the baker's yeast lineage. *PLoS Biol.* **2015**, *13*, e1002220. [[CrossRef](#)] [[PubMed](#)]
139. Albertin, W.; Marullo, P. Polyploidy in fungi: Evolution after whole-genome duplication. *Proc. Biol. Sci.* **2012**, *279*, 2497–2509. [[CrossRef](#)] [[PubMed](#)]
140. Aury, J.M. Global trends of whole-genome duplications revealed by the ciliate *Paramecium tetraurelia*. *Nature* **2006**, *444*, 171–178. [[CrossRef](#)] [[PubMed](#)]
141. De Bodt, S.; Maere, S.; van de Peer, Y. Genome duplication and the origin of angiosperms. *Trends Ecol. Evol.* **2005**, *20*, 591–597. [[CrossRef](#)] [[PubMed](#)]
142. Cui, L. Widespread genome duplications throughout the history of flowering plants. *Genome Res.* **2006**, *16*, 738–749. [[CrossRef](#)] [[PubMed](#)]
143. Dehal, P.; Boore, J.L. Two rounds of whole genome duplication in the ancestral vertebrate. *PLoS Biol.* **2005**, *3*. [[CrossRef](#)] [[PubMed](#)]
144. Kasahara, M.; Naruse, K.; Sasaki, S.; Nakatani, Y.; Qu, W.; Ahsan, B.; Yamada, T.; Nagayasu, Y.; Doi, K.; Kasai, Y.; *et al.* The medaka draft genome and insights into vertebrate genome evolution. *Nature* **2007**, *447*, 714–719. [[CrossRef](#)] [[PubMed](#)]
145. Kasahara, M. The 2R hypothesis: An update. *Curr. Opin. Immunol.* **2007**, *19*, 547–552. [[CrossRef](#)] [[PubMed](#)]
146. Donoghue, P.C.J.; Purnell, M.A. Genome duplication, extinction and vertebrate evolution. *Trends Ecol. Evol.* **2005**, *20*, 312–319. [[CrossRef](#)] [[PubMed](#)]
147. Hughes, T.; Liberles, D.A. Whole-genome duplications in the ancestral vertebrate are detectable in the distribution of gene family sizes of tetrapod species. *J. Mol. Evol.* **2008**, *67*, 343–357. [[CrossRef](#)] [[PubMed](#)]
148. Meyer, A.; van de Peer, Y. From 2R to 3R: Evidence for a fish-specific genome duplication (FSGD). *Bioessays* **2005**, *27*, 937–945. [[CrossRef](#)] [[PubMed](#)]
149. Glasauer, S.M.; Neuhauss, S.C. Whole-genome duplication in teleost fishes and its evolutionary consequences. *Mol. Genet. Genom.* **2014**, *289*, 1045–1060. [[CrossRef](#)] [[PubMed](#)]
150. Metcalfe, C.J.; Bulazel, K.V.; Ferreri, G.C.; Schroeder-Reiter, E.; Wanner, G.; Rens, W.; Obergefell, C.; Eldridge, M.D.; O'Neill, R.J. Genomic instability within centromeres of interspecific marsupial hybrids. *Genetics* **2007**, *177*, 2507–2517. [[CrossRef](#)] [[PubMed](#)]
151. Marfil, C.F.; Masuelli, R.W.; Davison, J.; Comai, L. Genomic instability in *Solanum tuberosum* × *Solanum kurtzianum* interspecific hybrids. *Genome* **2006**, *49*, 104–113. [[PubMed](#)]
152. Han, F.P.; Fedak, G.; Ouellet, T.; Liu, B. Rapid genomic changes in interspecific and intergeneric hybrids and allopolyploids of *Triticeae*. *Genome* **2003**, *46*, 716–723. [[CrossRef](#)] [[PubMed](#)]
153. King, M. *Species Evolution: The Role of Chromosome Change*; Cambridge University Press: Cambridge, UK, 1995.
154. White, M.J. Chromosomes of the vertebrates. *Evolution* **1949**, *3*, 379–381. [[CrossRef](#)] [[PubMed](#)]
155. Nie, W.; Wang, J.; Su, W.; Wang, D.; Tanomtung, A.; Perelman, P.L.; Graphodatsky, A.S.; Yang, F. Chromosomal rearrangements and karyotype evolution in carnivores revealed by chromosome painting. *Heredity* **2012**, *108*, 17–27. [[CrossRef](#)] [[PubMed](#)]
156. Lim, J.K.; Simmons, M.J. Gross chromosome rearrangements mediated by transposable elements in *Drosophila melanogaster*. *Bioessays* **1994**, *16*, 269–275. [[CrossRef](#)] [[PubMed](#)]
157. Mieczkowski, P.A.; Lemoine, F.J.; Petes, T.D. Recombination between retrotransposons as a source of chromosome rearrangements in the yeast *Saccharomyces cerevisiae*. *DNA Repair* **2006**, *5*, 1010–1020. [[CrossRef](#)] [[PubMed](#)]
158. Lonng, W.E.; Saedler, H. Chromosome rearrangements and transposable elements. *Ann. Rev. Genet.* **2002**, *36*, 389–410. [[CrossRef](#)] [[PubMed](#)]
159. Zhang, J.; Yu, C.; Krishnaswamy, L.; Peterson, T. Transposable elements as catalysts for chromosome rearrangements. *Methods Mol. Biol.* **2011**, *701*, 315–326. [[PubMed](#)]
160. Carbone, L.; Harris, R.A.; Gnerre, S.; Veeramah, K.R.; Lorente-Galdos, B.; Huddleston, J.; Meyer, T.J.; Herrero, J.; Roos, C.; Aken, B.; *et al.* Gibbon genome and the fast karyotype evolution of small apes. *Nature* **2014**, *513*, 195–201. [[CrossRef](#)] [[PubMed](#)]
161. Syvanen, M.; Kado, C.I. *Horizontal Gene Transfer*, 2nd ed.; Academic Press: London, UK, 2002.
162. Syvanen, M. Evolutionary implications of horizontal gene transfer. *Ann. Rev. Genet.* **2012**, *46*, 341–358. [[CrossRef](#)] [[PubMed](#)]
163. Watanabe, T. Infectious drug resistance. *Sci. Am.* **1967**, *217*, 19–28. [[CrossRef](#)] [[PubMed](#)]

164. Watanabe, T. Infective heredity of multiple drug resistance in bacteria. *Bacteriol. Rev.* **1963**, *27*, 87–115. [[PubMed](#)]
165. Andam, C.P.; Fournier, G.P.; Gogarten, J.P. Multilevel populations and the evolution of antibiotic resistance through horizontal gene transfer. *FEMS Microbiol. Rev.* **2011**, *35*, 756–767. [[CrossRef](#)] [[PubMed](#)]
166. Hastings, P.J.; Rosenberg, S.M.; Slack, A. Antibiotic-induced lateral transfer of antibiotic resistance. *Trends Microbiol.* **2004**, *12*, 401–404. [[CrossRef](#)] [[PubMed](#)]
167. Hayes, W. *The Genetics of Bacteria and Their Viruses*, 2nd ed.; Blackwell: London, UK, 1968.
168. *DNA Insertion Elements, Plasmids and Episomes*; Bukhari, A.I., Shapiro, J.A., Adhya, S.L., Eds.; Cold Spring Harbor Press: Cold Spring Harbor, New York, NY, USA, 1977.
169. Daccord, A.; Ceccarelli, D.; Rodrigue, S.; Burrus, V. Comparative analysis of mobilizable genomic islands. *J. Bacteriol.* **2013**, *195*, 606–614. [[CrossRef](#)] [[PubMed](#)]
170. Bellanger, X.; Payot, S.; Leblond-Bourget, N.; Guédon, G. Conjugative and mobilizable genomic islands in bacteria: Evolution and diversity. *FEMS Microbiol. Rev.* **2014**, *38*, 720–760. [[CrossRef](#)] [[PubMed](#)]
171. Makarova, K.S.; Wolf, Y.I.; Snir, S.; Koonin, E.V. Defense islands in bacterial and archaeal genomes and prediction of novel defense systems. *J. Bacteriol.* **2011**, *193*, 6039–6056. [[CrossRef](#)] [[PubMed](#)]
172. Van der Meer, J.R.; Sentchilo, V. Genomic islands and the evolution of catabolic pathways in bacteria. *Curr. Opin. Biotechnol.* **2003**, *14*, 248–254. [[CrossRef](#)]
173. Hall, R.M. Integrons and gene cassettes: Hotspots of diversity in bacterial genomes. *Ann. N.Y. Acad. Sci.* **2012**, *1267*, 71–78. [[CrossRef](#)] [[PubMed](#)]
174. Rowe-Magnus, D.A.; Mazel, D. The role of integrons in antibiotic resistance gene capture. *Int. J. Med. Microbiol.* **2002**, *292*, 115–125. [[CrossRef](#)] [[PubMed](#)]
175. Rowe-Magnus, D.A.; Guérout, A.M.; Mazel, D. Super-integrations. *Res. Microbiol.* **1999**, *150*, 641–651. [[CrossRef](#)]
176. Fluit, A.C.; Schmitz, F.J. Resistance integrons and super-integrations. *Clin. Microbiol. Infect.* **2004**, *10*, 272–288. [[CrossRef](#)] [[PubMed](#)]
177. Escudero, J.A.; Loot, C.; Nivina, A.; Mazel, D. The integron: Adaptation on demand. *Microbiol. Spectr.* **2015**, *3*. [[CrossRef](#)] [[PubMed](#)]
178. Rapa, R.A.; Labbate, M. The function of integron-associated gene cassettes in *Vibrio* species: The tip of the iceberg. *Front. Microbiol.* **2013**, *4*. [[CrossRef](#)] [[PubMed](#)]
179. Sclafani, R.A. Evidence for massive gene exchange between archaeal and bacterial hyperthermophiles. *Trends Genet.* **1998**, *14*, 442–444.
180. Nelson-Sathi, S.; Sousa, F.L.; Roettger, M.; Lozada-Chávez, N.; Thiery, T.; Janssen, A.; Bryant, D.; Landan, G.; Schönheit, P.; Siebers, B.; *et al.* Origins of major archaeal clades correspond to gene acquisitions from bacteria. *Nature* **2015**, *517*, 77–80. [[CrossRef](#)] [[PubMed](#)]
181. Dodsworth, J.A.; Li, L.; Wei, S.; Hedlund, B.P.; Leigh, J.A.; de Figueiredo, P. Inter-domain conjugal transfer of DNA from bacteria to archaea. *Appl. Environ. Microbiol.* **2010**, *76*, 5644–5647. [[CrossRef](#)] [[PubMed](#)]
182. Faguy, D.M.; Doolittle, W.F. Horizontal transfer of catalase-peroxidase genes between archaea and pathogenic bacteria. *Trends Genet.* **2000**, *16*, 196–197. [[CrossRef](#)]
183. Koonin, E.V.; Wolf, Y.I. Genomics of bacteria and archaea: The emerging dynamic view of the prokaryotic world. *Nucleic Acids Res.* **2008**, *36*, 6688–6719. [[CrossRef](#)] [[PubMed](#)]
184. Sonea, S.; Mathieu, L.G. Evolution of the genomic systems of prokaryotes and its momentous consequences. *Int. Microbiol.* **2001**, *4*, 67–71. [[PubMed](#)]
185. Sonea, S.; Panisset, M. *A New Bacteriology*; Jones and Batlett: Boston, MA, USA, 1983.
186. Sonea, S. A tentative unifying view of bacteria. *Rev. Can. Biol.* **1971**, *30*, 239–244. [[PubMed](#)]
187. Sharon, I.; Battchikova, N.; Aro, E.M.; Giglione, C.; Meinel, T.; Glaser, F.; Pinter, R.Y.; Breitbart, M.; Rohwer, F.; Béjà, O. Comparative metagenomics of microbial traits within oceanic viral communities. *ISME J.* **2011**, *5*, 1178–1190. [[CrossRef](#)] [[PubMed](#)]
188. Kristensen, D.M.; Mushegian, A.R.; Dolja, V.V.; Koonin, E.V. New dimensions of the virus world discovered through metagenomics. *Trends Microbiol.* **2010**, *18*, 11–19. [[CrossRef](#)] [[PubMed](#)]
189. Tamames, J.; Moya, A. Estimating the extent of horizontal gene transfer in metagenomic sequences. *BMC Genom.* **2008**, *9*. [[CrossRef](#)] [[PubMed](#)]
190. Ufarte, L.; Potocki-Veronese, G.; Laville, E. Discovery of new protein families and functions: New challenges in functional metagenomics for biotechnologies and microbial ecology. *Front. Microbiol.* **2015**, *6*. [[CrossRef](#)] [[PubMed](#)]



191. Wommack, K.E.; Nasko, D.J.; Chopyk, J.; Sakowski, E.G. Counts and sequences, observations that continue to change our understanding of viruses in nature. *J. Microbiol.* **2015**, *53*, 181–192. [[CrossRef](#)] [[PubMed](#)]
192. Labonté, J.M.; Field, E.K.; Lau, M.; Chivian, D.; van Heerden, E.; Wommack, K.E.; Kieft, T.L.; Onstott, T.C.; Stepanauskas, R. Single cell genomics indicates horizontal gene transfer and viral infections in a deep subsurface Firmicutes population. *Front. Microbiol.* **2015**, *6*. [[CrossRef](#)] [[PubMed](#)]
193. Mizuno, C.M.; Rodriguez-Valera, F.; Kimes, N.E.; Ghai, R. Expanding the marine virosphere using metagenomics. *PLoS Genet.* **2013**, *9*, e1003987. [[CrossRef](#)] [[PubMed](#)]
194. Lopez, P.; Halary, S.; Bapteste, E. Highly divergent ancient gene families in metagenomic samples are compatible with additional divisions of life. *Biol. Direct* **2015**, *10*. [[CrossRef](#)] [[PubMed](#)]
195. Moran, N.A.; Jarvik, T. Lateral transfer of genes from fungi underlies carotenoid production in aphids. *Science* **2010**, *328*, 624–627. [[CrossRef](#)] [[PubMed](#)]
196. Jackson, D.J.; Macis, L.; Reitner, J.; Wörheide, G. A horizontal gene transfer supported the evolution of an early metazoan biomineralization strategy. *BMC Evol. Biol.* **2011**, *11*. [[CrossRef](#)] [[PubMed](#)]
197. Altincicek, B.; Kovacs, J.L.; Gerardo, N.M. Horizontally transferred fungal carotenoid genes in the two-spotted spider mite *Tetranychus urticae*. *Biol. Lett.* **2012**, *8*, 253–257. [[CrossRef](#)] [[PubMed](#)]
198. Lane, N. Energetics and genetics across the prokaryote-eukaryote divide. *Biol. Direct* **2011**, *6*. [[CrossRef](#)] [[PubMed](#)]
199. Jaramillo, V.D.; Sukno, S.A.; Thon, M.R. Identification of horizontally transferred genes in the genus *Colletotrichum* reveals a steady tempo of bacterial to fungal gene transfer. *BMC Genom.* **2015**, *16*. [[CrossRef](#)] [[PubMed](#)]
200. Zhu, B.; Zhou, Q.; Xie, G.; Zhang, G.; Zhang, X.; Wang, Y.; Sun, G.; Li, B.; Jin, G. Interkingdom gene transfer may contribute to the evolution of phytopathogenicity in *Botrytis Cinerea*. *Evol. Bioinform. Online* **2012**, *8*, 105–117. [[CrossRef](#)] [[PubMed](#)]
201. Schönknecht, G.; Chen, W.H.; Ternes, C.M.; Barbier, G.G.; Shrestha, R.P.; Stanke, M.; Bräutigam, A.; Baker, B.J.; Banfield, J.F.; Garavito, R.M.; *et al.* Gene transfer from bacteria and archaea facilitated evolution of an extremophilic eukaryote. *Science* **2013**, *339*, 1207–1210. [[CrossRef](#)] [[PubMed](#)]
202. Bird, D.M.; Koltai, H. Plant parasitic nematodes: Habitats, hormones, and horizontally-acquired genes. *J. Plant Growth Regul.* **2000**, *19*, 183–194. [[PubMed](#)]
203. Baldwin, J.G.; Nadler, S.A.; Adams, B.J. Evolution of plant parasitism among nematodes. *Ann. Rev. Phytopathol.* **2004**, *42*, 83–105. [[CrossRef](#)] [[PubMed](#)]
204. Mitreva, M.; Smant, G.; Helder, J. Role of horizontal gene transfer in the evolution of plant parasitism among nematodes. *Methods Mol. Biol.* **2009**, *532*, 517–535. [[PubMed](#)]
205. Danchin, E.G.; Rosso, M.N.; Vieira, P.; de Almeida-Engler, J.; Coutinho, P.M.; Henrissat, B.; Abad, P. Multiple lateral gene transfers and duplications have promoted plant parasitism ability in nematodes. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 17651–17656. [[CrossRef](#)] [[PubMed](#)]
206. Haegeman, A.; Jones, J.T.; Danchin, E.G. Horizontal gene transfer in nematodes: A catalyst for plant parasitism? *Mol. Plant Microbe Interact.* **2011**, *24*, 879–887. [[CrossRef](#)] [[PubMed](#)]
207. Mayer, W.E.; Schuster, L.N.; Bartelmes, G.; Dieterich, C.; Sommer, R.J. Horizontal gene transfer of microbial cellulases into nematode genomes is associated with functional assimilation and gene turnover. *BMC Evol. Biol.* **2011**, *11*. [[CrossRef](#)] [[PubMed](#)]
208. Danchin, E.G.; Rosso, M.N. Lateral gene transfers have polished animal genomes: Lessons from nematodes. *Front. Cell. Infect. Microbiol.* **2012**, *2*. [[CrossRef](#)] [[PubMed](#)]
209. Yuan, J.B.; Zhang, X.J.; Liu, C.Z.; Wei, J.K.; Li, F.H.; Xiang, J.H. Horizontally transferred genes in the genome of Pacific white shrimp, *Litopenaeus vannamei*. *BMC Evol. Biol.* **2013**, *13*. [[CrossRef](#)] [[PubMed](#)]
210. Sun, B.F.; Li, T.; Xiao, J.H.; Jia, L.Y.; Liu, L.; Zhang, P.; Murphy, R.W.; He, S.M.; Huang, D.W. Horizontal functional gene transfer from bacteria to fishes. *Sci. Rep.* **2015**, *5*. [[CrossRef](#)] [[PubMed](#)]
211. Salzberg, S.L.; Hotopp, J.C.; Delcher, A.L.; Pop, M.; Smith, D.R.; Eisen, M.B.; Nelson, W.C. Serendipitous discovery of *Wolbachia* genomes in multiple *Drosophila* species. *Genome Biol.* **2005**, *6*. [[CrossRef](#)]
212. Hotopp, J.C.D.; Clark, M.E.; Oliveira, D.C.; Foster, J.M.; Fischer, P.; Torres, M.C.M.; Giebel, J.D.; Kumar, N.; Ishmael, N.; Wang, S.; *et al.* Widespread lateral gene transfer from intracellular bacteria to multicellular eukaryotes. *Science* **2007**, *317*, 1753–1756. [[CrossRef](#)] [[PubMed](#)]



213. Nikoh, N.; Tanaka, K.; Shibata, F.; Kondo, N.; Hizume, M.; Shimada, M.; Fukatsu, T. Wolbachia genome integrated in an insect chromosome: Evolution and fate of laterally transferred endosymbiont genes. *Genome Res.* **2008**, *18*, 272–280. [[CrossRef](#)] [[PubMed](#)]
214. Nikoh, N.; Nakabachi, A. Aphids acquired symbiotic genes via lateral gene transfer. *BMC Biol.* **2009**, *7*. [[CrossRef](#)] [[PubMed](#)]
215. Dunning Hotopp, J.C. Horizontal gene transfer between bacteria and animals. *Trends Genet.* **2011**, *27*, 157–163. [[CrossRef](#)] [[PubMed](#)]
216. Klasson, L.; Kumar, N.; Bromley, R.; Sieber, K.; Flowers, M.; Ott, S.H.; Tallon, L.J.; Andersson, S.G.; Dunning Hotopp, J.C. Extensive duplication of the *Wolbachia* DNA in chromosome four of *Drosophila ananassae*. *BMC Genom.* **2014**, *15*. [[CrossRef](#)] [[PubMed](#)]
217. Burstein, D.; Amaro, F.; Zusman, T.; Lifshitz, Z.; Cohen, O.; Gilbert, J.A.; Pupko, T.; Shuman, H.A.; Segal, G. Genomic analysis of 38 *Legionella* species identifies large and diverse effector repertoires. *Nat. Genet.* **2016**, *48*, 167–175. [[CrossRef](#)] [[PubMed](#)]
218. Bork, P. Hundreds of ankyrin-like repeats in functionally diverse proteins: Mobile modules that cross phyla horizontally? *Proteins* **1993**, *17*, 363–374. [[CrossRef](#)] [[PubMed](#)]
219. De Felipe, K.S.; Pampou, S.; Jovanovic, O.S.; Pericone, C.D.; Senna, F.Y.; Kalachikov, S.; Shuman, H.A. Evidence for acquisition of *Legionella* type IV secretion substrates via interdomain horizontal gene transfer. *J. Bacteriol.* **2005**, *187*, 7716–7726. [[CrossRef](#)] [[PubMed](#)]
220. De la Casa-Esperon, E. Horizontal transfer and the evolution of host-pathogen interactions. *Int. J. Evol. Biol.* **2012**, *2012*. [[CrossRef](#)] [[PubMed](#)]
221. Gomez-Valero, L.; Rusniok, C.; Jarraud, S.; Vacherie, B.; Rouy, Z.; Barbe, V.; Medigue, C.; Etienne, J.; Buchrieser, C. Extensive recombination events and horizontal gene transfer shaped the *Legionella pneumophila* genomes. *BMC Genom.* **2011**, *12*. [[CrossRef](#)] [[PubMed](#)]
222. Ensminger, A.W. *Legionella pneumophila*, armed to the hilt: Justifying the largest arsenal of effectors in the bacterial world. *Curr. Opin. Microbiol.* **2015**, *29*, 74–80. [[CrossRef](#)] [[PubMed](#)]
223. Jernigan, K.K.; Bordenstein, S.R. Ankyrin domains across the Tree of Life. *PeerJ* **2014**, *2*. [[CrossRef](#)] [[PubMed](#)]
224. Rennoll-Bankert, K.E.; Dumler, J.S. Lessons from *Anaplasma phagocytophilum*: Chromatin remodeling by bacterial effectors. *Infect. Disorders Drug Targets* **2012**, *12*, 380–387. [[CrossRef](#)]
225. Al-Khodori, S.; Price, C.T.; Kalia, A.; Kwaik, Y.A. Functional diversity of ankyrin repeats in microbial proteins. *Trends Microbiol.* **2010**, *18*, 132–139. [[CrossRef](#)] [[PubMed](#)]
226. Habyarimana, F.; Price, C.T.; Santic, M.; Al-Khodori, S.; Kwaik, Y.A. Molecular characterization of the Dot/Icm-translocated AnkH and AnkJ eukaryotic-like effectors of *Legionella pneumophila*. *Infect. Immun.* **2010**, *78*, 1123–1134. [[CrossRef](#)] [[PubMed](#)]
227. Voth, D.E. ThANKs for the repeat: Intracellular pathogens exploit a common eukaryotic domain. *Cell. Logist.* **2011**, *1*, 128–132. [[CrossRef](#)] [[PubMed](#)]
228. Dubreuil, R.; Segev, N. Bringing host-cell takeover by pathogenic bacteria to center stage. *Cell. Logist.* **2011**, *1*, 120–124. [[CrossRef](#)] [[PubMed](#)]
229. Gomez-Valero, L.; Rusniok, C.; Cazalet, C.; Buchrieser, C. Comparative and functional genomics of legionella identified eukaryotic like proteins as key players in host-pathogen interactions. *Front. Microbiol.* **2011**, *2*. [[CrossRef](#)] [[PubMed](#)]
230. Gomez-Valero, L.; Buchrieser, C. Genome dynamics in *Legionella*: The basis of versatility and adaptation to intracellular replication. *Cold Spring Harb. Perspect. Med.* **2013**, *3*. [[CrossRef](#)] [[PubMed](#)]
231. Keeling, P.J.; Palmer, J.D. Horizontal gene transfer in eukaryotic evolution. *Nat. Rev. Genet.* **2008**, *9*, 605–618. [[CrossRef](#)] [[PubMed](#)]
232. Andersson, J.O.; Sjögren, Å.M.; Davis, L.A.; Embley, T.M.; Roger, A.J. Phylogenetic analyses of diplomonad genes reveal frequent lateral gene transfers affecting eukaryotes. *Curr. Biol.* **2003**, *13*, 94–104. [[CrossRef](#)]
233. Alsmark, U.C.; Sicheritz-Ponten, T.; Foster, P.G.; Hirt, R.P.; Embley, T.M. Horizontal gene transfer in eukaryotic parasites: A case study of *Entamoeba histolytica* and *Trichomonas vaginalis*. *Methods Mol. Biol.* **2009**, *532*, 489–500. [[PubMed](#)]
234. Alsmark, C.; Foster, P.G.; Sicheritz-Ponten, T.; Nakjang, S.; Embley, T.M.; Hirt, R.P. Patterns of prokaryotic lateral gene transfers affecting parasitic microbial eukaryotes. *Genome Biol.* **2013**, *14*. [[CrossRef](#)] [[PubMed](#)]
235. Xi, Z.; Wang, Y.; Bradley, R.K.; Sugumaran, M.; Marx, C.J.; Rest, J.S.; Davis, C.C. Massive mitochondrial gene transfer in a parasitic flowering plant clade. *PLoS Genet.* **2013**, *9*, e1003265. [[CrossRef](#)] [[PubMed](#)]

236. Hao, W.; Richardson, A.O.; Zheng, Y.; Palmer, J.D. Gorgeous mosaic of mitochondrial genes created by horizontal transfer and gene conversion. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 21576–21581. [[CrossRef](#)] [[PubMed](#)]
237. Bergthorsson, U.; Adams, K.L.; Thomason, B.; Palmer, J.D. Widespread horizontal transfer of mitochondrial genes in flowering plants. *Nature* **2003**, *424*, 197–201. [[CrossRef](#)] [[PubMed](#)]
238. Stegemann, S.; Keuthe, M.; Greiner, S.; Bock, R. Horizontal transfer of chloroplast genomes between plant species. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 2434–2438. [[CrossRef](#)] [[PubMed](#)]
239. Pace, J.K.; Gilbert, C.; Clark, M.S.; Feschotte, C. Repeated horizontal transfer of a DNA transposon in mammals and other tetrapods. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17023–17028. [[CrossRef](#)] [[PubMed](#)]
240. Fortune, P.M.; Roulin, A.; Panaud, O. Horizontal transfer of transposable elements in plants. *Commun. Integr. Biol.* **2008**, *1*, 74–77. [[CrossRef](#)] [[PubMed](#)]
241. Thomas, J.; Schaack, S.; Pritham, E.J. Pervasive horizontal transfer of rolling-circle transposons among animals. *Genome Biol. Evol.* **2010**, *2*, 656–664. [[CrossRef](#)] [[PubMed](#)]
242. Bartolome, C.; Bello, X.; Maside, X. Widespread evidence for horizontal transfer of transposable elements across *Drosophila* genomes. *Genome Biol.* **2009**, *10*. [[CrossRef](#)] [[PubMed](#)]
243. Novick, P.; Smith, J.; Ray, D.; Boissinot, S. Independent and parallel lateral transfer of DNA transposons in tetrapod genomes. *Gene* **2010**, *449*, 85–94. [[CrossRef](#)] [[PubMed](#)]
244. Wallau, G.L.; Ortiz, M.F.; Loreto, E.L. Horizontal transposon transfer in eukarya: Detection, bias, and perspectives. *Genome Biol. Evol.* **2012**, *4*, 689–699. [[CrossRef](#)] [[PubMed](#)]
245. Ivancevic, A.M.; Walsh, A.M.; Kortschak, R.D.; Adelson, D.L. Jumping the fine LINE between species: Horizontal transfer of transposable elements in animals catalyses genome evolution. *Bioessays* **2013**, *35*, 1071–1082. [[CrossRef](#)] [[PubMed](#)]
246. Kondrashov, F.A.; Koonin, E.V.; Morgunov, I.G.; Finogenova, T.V.; Kondrashova, M.N. Evolution of glyoxylate cycle enzymes in Metazoa: Evidence of multiple horizontal transfer events and pseudogene formation. *Biol. Direct* **2006**, *1*. [[CrossRef](#)] [[PubMed](#)]
247. Rogers, M.; Keeling, P.J. Lateral transfer and re compartmentalization of Calvin cycle enzymes of plants and algae. *J. Mol. Evol.* **2004**, *58*, 367–375. [[CrossRef](#)] [[PubMed](#)]
248. Christin, P.A.; Wallace, M.J.; Clayton, H.; Edwards, E.J.; Furbank, R.T.; Hattersley, P.W.; Sage, R.F.; Macfarlane, T.D.; Ludwig, M. Multiple photosynthetic transitions, polyploidy, and lateral gene transfer in the grass subtribe *Neurachninae*. *J. Exp. Bot.* **2012**, *63*, 6297–6308. [[CrossRef](#)] [[PubMed](#)]
249. Graham, L.A.; Loughheed, S.C.; Ewart, K.V.; Davies, P.L. Lateral transfer of a lectin-like antifreeze protein gene in fishes. *PLoS ONE* **2008**, *3*, e2616. [[CrossRef](#)] [[PubMed](#)]
250. Heliconius Genome Consortium. Butterfly genome reveals promiscuous exchange of mimicry adaptations among species. *Nature* **2012**, *487*, 94–98.
251. Xi, Z.; Bradley, R.K.; Wurdack, K.J.; Wong, K.; Sugumaran, M.; Bomblies, K.; Rest, J.S.; Davis, C.C. Horizontal transfer of expressed genes in a parasitic flowering plant. *BMC Genom.* **2012**, *13*. [[CrossRef](#)] [[PubMed](#)]
252. Houck, M.A.; Clark, J.B.; Peterson, K.R.; Kidwell, M.G. Possible horizontal transfer of *Drosophila* genes by the mite *Proctolaelaps regalis*. *Science* **1991**, *253*, 1125–1128. [[CrossRef](#)] [[PubMed](#)]
253. Gilbert, C.; Schaack, S.; Pace, J.K., II; Brindley, P.J.; Feschotte, C. A role for host-parasite interactions in the horizontal transfer of transposons across phyla. *Nature* **2010**, *464*, 1347–1350. [[CrossRef](#)] [[PubMed](#)]
254. Barteneva, N.S.; Maltsev, N.; Vorobjev, I.A. Microvesicles and intercellular communication in the context of parasitism. *Front. Cell. Infect. Microbiol.* **2013**, *3*. [[CrossRef](#)] [[PubMed](#)]
255. Qiu, H.; Yoon, H.S.; Bhattacharya, D. Algal endosymbionts as vectors of horizontal gene transfer in photosynthetic eukaryotes. *Front. Plant Sci.* **2013**, *4*. [[CrossRef](#)] [[PubMed](#)]
256. Taylor, M.; Mediannikov, O.; Raoult, D.; Greub, G. Endosymbiotic bacteria associated with nematodes, ticks and amoebae. *FEMS Immunol. Med. Microbiol.* **2012**, *64*, 21–31. [[CrossRef](#)] [[PubMed](#)]
257. Sandström, J.P.; Russell, J.A.; White, J.P.; Moran, N.A. Independent origins and horizontal transfer of bacterial symbionts of aphids. *Mol. Ecol.* **2001**, *10*, 217–228. [[CrossRef](#)] [[PubMed](#)]
258. Raychoudhury, R.; Baldo, L.; Oliveira, D.C.; Werren, J.H. Modes of acquisition of *Wolbachia*: Horizontal transfer, hybrid introgression, and codivergence in the *Nasonia* species complex. *Evolution* **2009**, *63*, 165–183. [[CrossRef](#)] [[PubMed](#)]

259. Oliver, K.M.; Degnan, P.H.; Burke, G.R.; Moran, N.A. Facultative symbionts in aphids and the horizontal transfer of ecologically important traits. *Ann. Rev. Entomol.* **2010**, *55*, 247–266. [[CrossRef](#)] [[PubMed](#)]
260. Bozzaro, S.; Eichinger, L. The professional phagocyte *Dictyostelium discoideum* as a model host for bacterial pathogens. *Curr. Drug Targets* **2011**, *12*, 942–954. [[CrossRef](#)] [[PubMed](#)]
261. Chien, M.; Morozova, I.; Shi, S.; Sheng, H.; Chen, J.; Gomez, S.M.; Asamani, G.; Hill, K.; Nuara, J.; Feder, M.; et al. The genomic sequence of the accidental pathogen *Legionella pneumophila*. *Science* **2004**, *305*, 1966–1968. [[CrossRef](#)] [[PubMed](#)]
262. Steinert, M. Pathogen-host interactions in *Dictyostelium*, *Legionella*, *Mycobacterium* and other pathogens. *Semin. Cell Dev. Biol.* **2011**, *22*, 70–76. [[CrossRef](#)] [[PubMed](#)]
263. Huws, S.A.; Morley, R.J.; Jones, M.V.; Brown, M.R.; Smith, A.W. Interactions of some common pathogenic bacteria with *Acanthamoeba polyphaga*. *FEMS Microbiol. Lett.* **2008**, *282*, 258–265. [[CrossRef](#)] [[PubMed](#)]
264. Douesnard-Malo, F.; Daigle, F. Increased persistence of *Salmonella enterica* serovar Typhi in the presence of *Acanthamoeba castellanii*. *Appl. Environ. Microbiol.* **2011**, *77*, 7640–7646. [[CrossRef](#)] [[PubMed](#)]
265. Yousuf, F.A.; Siddiqui, R.; Khan, N.A. *Acanthamoeba castellanii* of the T4 genotype is a potential environmental host for *Enterobacter aerogenes* and *Aeromonas hydrophila*. *Parasites Vectors* **2013**, *6*. [[CrossRef](#)] [[PubMed](#)]
266. Jeon, K.W. Genetic and physiological interactions in the amoeba-bacteria symbiosis. *J. Eukaryot. Microbiol.* **2004**, *51*, 502–508. [[CrossRef](#)] [[PubMed](#)]
267. Charpentier, X.; Kay, E.; Schneider, D.; Shuman, H.A. Antibiotics and UV radiation induce competence for natural transformation in *Legionella pneumophila*. *J. Bacteriol.* **2011**, *193*, 1114–1121. [[CrossRef](#)] [[PubMed](#)]
268. Sun, Y.; Bernardy, E.E.; Hammer, B.K.; Miyashiro, T. Competence and natural transformation in vibrios. *Mol. Microbiol.* **2013**, *89*, 583–595. [[CrossRef](#)] [[PubMed](#)]
269. Kovács, Á.T.; Smits, W.K.; Mirończuk, A.M.; Kuipers, O.P. Ubiquitous late competence genes in *Bacillus* species indicate the presence of functional DNA uptake machineries. *Environ. Microbiol.* **2009**, *11*, 1911–1922. [[CrossRef](#)] [[PubMed](#)]
270. Benam, A.V.; Lång, E.; Alfsnes, K.; Fleckenstein, B.; Rowe, A.D.; Hovland, E.; Ambur, O.H.; Frye, S.A.; Tønjum, T. Structure-function relationships of the competence lipoprotein ComL and SSB in meningococcal transformation. *Microbiology* **2011**, *157*, 1329–1342. [[CrossRef](#)] [[PubMed](#)]
271. Crochu, S.; Cook, S.; Attoui, H.; Charrel, R.N.; de Chesse, R.; Belhouchet, M.; Lemasson, J.J.; de Micco, P.; de Lamballerie, X. Sequences of flavivirus-related RNA viruses persist in DNA form integrated in the genome of *Aedes spp.* mosquitoes. *J. Gen. Virol.* **2004**, *85*, 1971–1980. [[CrossRef](#)] [[PubMed](#)]
272. Tanne, E.; Sela, I. Occurrence of a DNA sequence of a non-retro RNA virus in a host plant genome and its expression: Evidence for recombination between viral and host RNAs. *Virology* **2005**, *332*, 614–622. [[CrossRef](#)] [[PubMed](#)]
273. Frank, A.C.; Wolfe, K.H. Evolutionary capture of viral and plasmid DNA by yeast nuclear chromosomes. *Eukaryot. Cell* **2009**, *8*, 1521–1531. [[CrossRef](#)] [[PubMed](#)]
274. Roiz, D.; Vázquez, A.; Seco, M.P.S.; Tenorio, A.; Rizzoli, A. Detection of novel insect flavivirus sequences integrated in *Aedes albopictus* (Diptera: Culicidae) in Northern Italy. *Virol. J.* **2009**, *6*. [[CrossRef](#)] [[PubMed](#)]
275. Taylor, D.J.; Leach, R.W.; Bruenn, J. Filoviruses are ancient and integrated into mammalian genomes. *BMC Evol. Biol.* **2010**, *10*. [[CrossRef](#)] [[PubMed](#)]
276. Belyi, V.A.; Levine, A.J.; Skalka, A.M. Sequences from ancestral single-stranded DNA viruses in vertebrate genomes: The *Parvoviridae* and *Circoviridae* are more than 40 to 50 million years old. *J. Virol.* **2010**, *84*, 12458–12462. [[CrossRef](#)] [[PubMed](#)]
277. Belyi, V.A.; Levine, A.J.; Skalka, A.M. Unexpected inheritance: Multiple integrations of ancient bornavirus and ebolavirus/marburgvirus sequences in vertebrate genomes. *PLoS Pathog.* **2010**, *6*, e1001030. [[CrossRef](#)] [[PubMed](#)]
278. Horie, M.; Honda, T.; Suzuki, Y.; Kobayashi, Y.; Daito, T.; Oshida, T.; Ikuta, K.; Jern, P.; Gojobori, T.; Coffin, J.M.; et al. Endogenous non-retroviral RNA virus elements in mammalian genomes. *Nature* **2010**, *463*, 84–87. [[CrossRef](#)] [[PubMed](#)]
279. Iskra-Caruana, M.L.; Baurens, F.C.; Gayral, P.; Chabannes, M. A four-partner plant-virus interaction: Enemies can also come from within. *Mol. Plant Microbe Interact.* **2010**, *23*, 1394–1402. [[CrossRef](#)] [[PubMed](#)]
280. Kapoor, A.; Simmonds, P.; Lipkin, W.I. Discovery and characterization of mammalian endogenous parvoviruses. *J. Virol.* **2010**, *84*, 12628–12635. [[CrossRef](#)] [[PubMed](#)]

281. Katzourakis, A.; Gifford, R.J. Endogenous viral elements in animal genomes. *PLoS Genet.* **2010**, *6*, e1001191. [[CrossRef](#)] [[PubMed](#)]
282. Liu, H.; Fu, Y.; Jiang, D.; Li, G.; Xie, J.; Cheng, J.; Peng, Y.; Ghabrial, S.A.; Yi, X. Widespread horizontal gene transfer from double-stranded RNA viruses to eukaryotic nuclear genomes. *J. Virol.* **2010**, *84*, 11876–11887. [[CrossRef](#)] [[PubMed](#)]
283. Horie, M.; Tomonaga, K. Non-retroviral fossils in vertebrate genomes. *Viruses* **2011**, *3*, 1836–1848. [[CrossRef](#)] [[PubMed](#)]
284. Chiba, S.; Kondo, H.; Tani, A.; Saisho, D.; Sakamoto, W.; Kanematsu, S.; Suzuki, N. Widespread endogenization of genome sequences of non-retroviral RNA viruses into plant genomes. *PLoS Pathog.* **2011**, *7*, e1002146. [[CrossRef](#)] [[PubMed](#)]
285. Liu, H.; Fu, Y.; Xie, J.; Cheng, J.; Ghabrial, S.A.; Li, G.; Peng, Y.; Yi, X.; Jiang, D. Widespread endogenization of densoviruses and parvoviruses in animal and human genomes. *J. Virol.* **2011**, *85*, 9863–9876. [[CrossRef](#)] [[PubMed](#)]
286. Holmes, E.C. The evolution of endogenous viral elements. *Cell Host Microbe* **2011**, *10*, 368–377. [[CrossRef](#)] [[PubMed](#)]
287. Feschotte, C.; Gilbert, C. Endogenous viruses: Insights into viral evolution and impact on host biology. *Nat. Rev. Genet.* **2012**, *13*, 283–296. [[CrossRef](#)] [[PubMed](#)]
288. Cui, J.; Holmes, E.C. Endogenous RNA viruses of plants in insect genomes. *Virology* **2012**, *427*, 77–79. [[CrossRef](#)] [[PubMed](#)]
289. Geuking, M.B.; Weber, J.; Dewannieux, M.; Gorelik, E.; Heidmann, T.; Hengartner, H.; Zinkernagel, R.M.; Hangartner, L. Recombination of retrotransposon and exogenous RNA virus results in nonretroviral cDNA integration. *Science* **2009**, *323*, 393–396. [[CrossRef](#)] [[PubMed](#)]
290. Bill, C.A.; Summers, J. Genomic DNA double-strand breaks are targets for hepadnaviral DNA integration. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 11135–11140. [[CrossRef](#)] [[PubMed](#)]
291. Hu, X.; Lin, J.; Xie, Q.; Ren, J.; Chang, Y.; Wu, W.; Xia, Y. DNA double-strand breaks, potential targets for HBV integration. *J. Huazhong Univ. Sci. Technol. Med. Sci.* **2010**, *30*, 265–270. [[CrossRef](#)] [[PubMed](#)]
292. Koonin, E.V.; Dolja, V.V. Virus world as an evolutionary network of viruses and capsidless selfish elements. *Microbiol. Mol. Biol. Rev.* **2014**, *78*, 278–303. [[CrossRef](#)] [[PubMed](#)]
293. Abroi, A. A protein domain-based view of the virosphere-host relationship. *Biochimie* **2015**, *119*, 231–243. [[CrossRef](#)] [[PubMed](#)]
294. Yoosuf, N.; Yutin, N.; Colson, P.; Shabalina, S.A.; Pagnier, I.; Robert, C.; Azza, S.; Klose, T.; Wong, J.; Rossmann, M.G.; *et al.* Related giant viruses in distant locations and different habitats: *Acanthamoeba polyphaga* moomouvirus represents a third lineage of the *Mimiviridae* that is close to the megavirus lineage. *Genome Biol. Evol.* **2012**, *4*, 1324–1330. [[CrossRef](#)] [[PubMed](#)]
295. Claverie, J.-M.; Abergel, C. The concept of virus in the post-megavirus era. In *Viruses: Essential Agents of Life*; Witzany, G., Ed.; Springer: Dordrecht, The Netherlands, 2012; pp. 187–202.
296. Piacente, F.; de Castro, C.; Jeudy, S.; Molinaro, A.; Salis, A.; Damonte, G.; Bernardi, C.; Abergel, C.; Tonetti, M.G. Giant virus *Megavirus chilensis* encodes the biosynthetic pathway for uncommon acetamido sugars. *J. Biol. Chem.* **2014**, *289*, 24428–24439. [[CrossRef](#)] [[PubMed](#)]
297. Filee, J.; Pouget, N.; Chandler, M. Phylogenetic evidence for extensive lateral acquisition of cellular genes by Nucleocytoplasmic large DNA viruses. *BMC Evol. Biol.* **2008**, *8*. [[CrossRef](#)] [[PubMed](#)]
298. Filee, J. Lateral gene transfer, lineage-specific gene expansion and the evolution of Nucleo Cytoplasmic Large DNA viruses. *J. Invertebrate Pathol.* **2009**, *101*, 169–171. [[CrossRef](#)] [[PubMed](#)]
299. Filee, J.; Chandler, M. Gene exchange and the origin of giant viruses. *Intervirology* **2010**, *53*, 354–361. [[CrossRef](#)] [[PubMed](#)]
300. Filee, J. Route of NCLDV evolution: The genomic accordion. *Curr. Opin. Virol.* **2013**, *3*, 595–599. [[CrossRef](#)] [[PubMed](#)]
301. Boyer, M.; Yutin, N.; Pagnier, I.; Barrassi, L.; Fournous, G.; Espinosa, L.; Robert, C.; Azza, S.; Sun, S.; Rossmann, M.G.; *et al.* Giant Marseillevirus highlights the role of *Amoebae* as a melting pot in emergence of chimeric microorganisms. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 21848–21853. [[CrossRef](#)] [[PubMed](#)]
302. Colson, P.; Gimenez, G.; Boyer, M.; Fournous, G.; Raoult, D. The giant *Cafeteria roenbergensis* virus that infects a widespread marine phagocytic protist is a new member of the fourth domain of life. *PLoS ONE* **2011**, *6*, e18935. [[CrossRef](#)] [[PubMed](#)]



303. Saisongkorh, W.; Robert, C.; la Scola, B.; Raoult, D.; Rolain, J.M. Evidence of transfer by conjugation of type IV secretion system genes between *Bartonella* species and *Rhizobium radiobacter* in amoeba. *PLoS ONE* **2010**, *5*, e12666. [[CrossRef](#)] [[PubMed](#)]
304. McClure, M.A. Evolution of the DUT gene: Horizontal transfer between host and pathogen in all three domains of life. *Curr. Protein Pept. Sci.* **2001**, *2*, 313–324. [[CrossRef](#)] [[PubMed](#)]
305. Metcalf, J.A.; Funkhouser-Jones, L.J.; Briley, K.; Reysenbach, A.L.; Bordenstein, S.R. Antibacterial gene transfer across the tree of life. *eLife* **2014**, *3*. [[CrossRef](#)] [[PubMed](#)]
306. Doolittle, R.F.; Bork, P. Evolutionarily mobile modules in proteins. *Sci. Am.* **1993**, *269*, 50–56. [[CrossRef](#)] [[PubMed](#)]
307. Lander, E.S.; Linton, L.M.; Birren, B.; Nusbaum, C.; Zody, M.C.; Baldwin, J.; Devon, K.; Dewar, K.; Doyle, M.; FitzHugh, W.; *et al.* Initial sequencing and analysis of the human genome. *Nature* **2001**, *409*, 860–921. [[CrossRef](#)] [[PubMed](#)]
308. Schmitz, J.; Brosius, J. Exonization of transposed elements: A challenge and opportunity for evolution. *Biochimie* **2011**, *93*, 1928–1934. [[CrossRef](#)] [[PubMed](#)]
309. Gilbert, W. The exon theory of genes. *Cold Spring Harb. Symp. Quant. Biol.* **1987**, *52*, 901–905. [[CrossRef](#)] [[PubMed](#)]
310. Gilbert, W. DNA sequencing and gene structure. *Science* **1981**, *214*, 1305–1312. [[CrossRef](#)] [[PubMed](#)]
311. Liu, M.; Grigoriev, A. Protein domains correlate strongly with exons in multiple eukaryotic genomes—Evidence of exon shuffling? *Trends Genet.* **2004**, *20*, 399–403. [[CrossRef](#)] [[PubMed](#)]
312. Xing, Y.; Lee, C. Alternative splicing and RNA selection pressure—Evolutionary consequences for eukaryotic genomes. *Nat. Rev. Genet.* **2006**, *7*, 499–509. [[CrossRef](#)] [[PubMed](#)]
313. Barbosa-Morais, N.L.; Irimia, M.; Pan, Q.; Xiong, H.Y.; Gueroussov, S.; Lee, L.J.; Slobodeniuc, V.; Kutter, C.; Watt, S.; Çolak, R.; *et al.* The evolutionary landscape of alternative splicing in vertebrate species. *Science* **2012**, *338*, 1587–1593. [[CrossRef](#)] [[PubMed](#)]
314. Hassan, M.A.; Saeij, J.P. Incorporating alternative splicing and mRNA editing into the genetic analysis of complex traits. *Bioessays* **2014**, *36*, 1032–1040. [[CrossRef](#)] [[PubMed](#)]
315. Kornblihtt, A.R.; Schor, I.E.; Alló, M.; Dujardin, G.; Petrillo, E.; Muñoz, M.J. Alternative splicing: A pivotal step between eukaryotic transcription and translation. *Nat. Rev. Mol. Cell Biol.* **2013**, *14*, 153–165. [[CrossRef](#)] [[PubMed](#)]
316. Kalsotra, A.; Cooper, T.A. Functional consequences of developmentally regulated alternative splicing. *Nat. Rev. Genet.* **2011**, *12*, 715–729. [[CrossRef](#)] [[PubMed](#)]
317. Ast, G. How did alternative splicing evolve? *Nat. Rev. Genet.* **2004**, *5*, 773–782. [[CrossRef](#)] [[PubMed](#)]
318. Chen, M.; Manley, J.L. Mechanisms of alternative splicing regulation: Insights from molecular and genomics approaches. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 741–754. [[CrossRef](#)] [[PubMed](#)]
319. Mudge, J.M.; Frankish, A.; Fernandez-Banet, J.; Alioto, T.; Derrien, T.; Howald, C.; Reymond, A.; Guigó, R.; Hubbard, T.; Harrow, J. The origins, evolution, and functional potential of alternative splicing in vertebrates. *Mol. Biol. Evol.* **2011**, *28*, 2949–2959. [[CrossRef](#)] [[PubMed](#)]
320. Kawashima, T.; Kawashima, S.; Tanaka, C.; Murai, M.; Yoneda, M.; Putnam, N.H.; Rokhsar, D.S.; Kanehisa, M.; Satoh, N.; Wada, H. Domain shuffling and the evolution of vertebrates. *Genome Res.* **2009**, *19*, 1393–1403. [[CrossRef](#)] [[PubMed](#)]
321. Kaessmann, H.; Zöllner, S.; Nekrutenko, A.; Li, W.H. Signatures of domain shuffling in the human genome. *Genome Res.* **2002**, *12*, 1642–1650. [[CrossRef](#)] [[PubMed](#)]
322. Van Rijk, A.; Bloemendal, H. Molecular mechanisms of exon shuffling: Illegitimate recombination. *Genetica* **2003**, *118*, 245–249. [[CrossRef](#)] [[PubMed](#)]
323. Franca, G.S.; Cancherini, D.V.; de Souza, S.J. Evolutionary history of exon shuffling. *Genetica* **2012**, *140*, 249–257. [[CrossRef](#)] [[PubMed](#)]
324. Sorek, R. The birth of new exons: Mechanisms and evolutionary consequences. *RNA* **2007**, *13*, 1603–1608. [[CrossRef](#)] [[PubMed](#)]
325. Liu, L.Y.; Charng, Y.C. Genome-wide survey of ds exonization to enrich transcriptomes and proteomes in plants. *Evolut. Bioinform. Online* **2012**, *8*, 575–587.
326. Huda, A.; Bushel, P.R. Widespread exonization of transposable elements in human coding sequences is associated with epigenetic regulation of transcription. *Transcriptomics Open Access* **2013**, *1*. [[CrossRef](#)]



327. Bacher, J.M.; Reiss, B.D.; Ellington, A.D. Anticipatory evolution and DNA shuffling. *Genome Biol.* **2002**, *3*. [[CrossRef](#)]
328. Stemmer, W.P. DNA shuffling by random fragmentation and reassembly: *In vitro* recombination for molecular evolution. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 10747–10751. [[CrossRef](#)] [[PubMed](#)]
329. Stemmer, W.P. Rapid evolution of a protein *in vitro* by DNA shuffling. *Nature* **1994**, *370*, 389–391. [[CrossRef](#)] [[PubMed](#)]
330. Ejima, Y.; Yang, L. Trans mobilization of genomic DNA as a mechanism for retrotransposon-mediated exon shuffling. *Hum. Mol. Genet.* **2003**, *12*, 1321–1328. [[CrossRef](#)] [[PubMed](#)]
331. Jiang, N.; Bao, Z.; Zhang, X.; Eddy, S.R.; Wessler, S.R. Pack-MULE transposable elements mediate gene evolution in plants. *Nature* **2004**, *431*, 569–573. [[CrossRef](#)] [[PubMed](#)]
332. Morgante, M.; Brunner, S.; Pea, G.; Fengler, K.; Zuccolo, A.; Rafalski, A. Gene duplication and exon shuffling by helitron-like transposons generate intraspecies diversity in maize. *Nat. Genet.* **2005**, *37*, 997–1002. [[CrossRef](#)] [[PubMed](#)]
333. Lisch, D. Pack-MULEs: Theft on a massive scale. *Bioessays* **2005**, *27*, 353–355. [[CrossRef](#)] [[PubMed](#)]
334. Damert, A.; Raiz, J.; Horn, A.V.; Löwer, J.; Wang, H.; Xing, J.; Batzer, M.A.; Löwer, R.; Schumann, G.G. 5'-Transducing SVA retrotransposon groups spread efficiently throughout the human genome. *Genome Res.* **2009**, *19*, 1992–2008. [[CrossRef](#)] [[PubMed](#)]
335. Hanks, D.C.; Ewing, A.D.; Chen, J.E.; Tokunaga, K.; Kazazian, H.H., Jr. Exon-trapping mediated by the human retrotransposon SVA. *Genome Res.* **2009**, *19*, 1983–1991. [[CrossRef](#)] [[PubMed](#)]
336. Elrouby, N.; Bureau, T.E. *Bs1*, a new chimeric gene formed by retrotransposon-mediated exon shuffling in maize. *Plant Physiol.* **2010**, *153*, 1413–1424. [[CrossRef](#)] [[PubMed](#)]
337. Jiang, N.; Ferguson, A.A.; Slotkin, R.K.; Lisch, D. Pack-Mutator-like transposable elements (Pack-MULEs) induce directional modification of genes through biased insertion and DNA acquisition. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 1537–1542. [[CrossRef](#)] [[PubMed](#)]
338. Moran, J.V.; DeBerardinis, R.J.; Kazazian, H.H., Jr. Exon shuffling by L1 retrotransposition. *Science* **1999**, *283*, 1530–1534. [[CrossRef](#)] [[PubMed](#)]
339. Hiller, R.; Hetzer, M.; Schweyen, R.J.; Mueller, M.W. Transposition and exon shuffling by group II intron RNA molecules in pieces. *J. Mol. Biol.* **2000**, *297*, 301–308. [[CrossRef](#)] [[PubMed](#)]
340. Kolkman, J.A.; Stemmer, W.P. Directed evolution of proteins by exon shuffling. *Nat. Biotechnol.* **2001**, *19*, 423–428. [[CrossRef](#)] [[PubMed](#)]
341. Piriyaopongsa, J.; Polavarapu, N.; Borodovsky, M.; McDonald, J. Exonization of the LTR transposable elements in human genome. *BMC Genom.* **2007**, *8*. [[CrossRef](#)] [[PubMed](#)]
342. Schwartz, S.; Gal-Mark, N.; Kfir, N.; Oren, R.; Kim, E.; Ast, G. Alu exonization events reveal features required for precise recognition of exons by the splicing machinery. *PLoS Comput. Biol.* **2009**, *5*, e1000300. [[CrossRef](#)] [[PubMed](#)]
343. Sela, N.; Mersch, B.; Hotz-Wagenblatt, A.; Ast, G. Characteristics of transposable element exonization within human and mouse. *PLoS ONE* **2010**, *5*, e10907. [[CrossRef](#)] [[PubMed](#)]
344. Krull, M.; Brosius, J.; Schmitz, J. Alu-SINE exonization: En route to protein-coding function. *Mol. Biol. Evol.* **2005**, *22*, 1702–1711. [[CrossRef](#)] [[PubMed](#)]
345. Möller-Krull, M.; Zemmann, A.; Roos, C.; Brosius, J.; Schmitz, J. Beyond DNA: RNA editing and steps toward Alu exonization in primates. *J. Mol. Biol.* **2008**, *382*, 601–609. [[CrossRef](#)] [[PubMed](#)]
346. Zemojtel, T.; Penzkofer, T.; Schultz, J.; Dandekar, T.; Badge, R.; Vingron, M. Exonization of active mouse L1s: A driver of transcriptome evolution? *BMC Genom.* **2007**, *8*. [[CrossRef](#)] [[PubMed](#)]
347. Dobzhansky, T. *Genetics and the Origin of Species*; Columbia University Press: New York, NY, USA, 1937.
348. Marino-Ramirez, L.; Lewis, K.C.; Landsman, D.; Jordan, I.K. Transposable elements donate lineage-specific regulatory sequences to host genomes. *Cytogenetic Genome Res.* **2005**, *110*, 333–341. [[CrossRef](#)] [[PubMed](#)]
349. Wang, T.; Zeng, J.; Lowe, C.B.; Sellers, R.G.; Salama, S.R.; Yang, M.; Burgess, S.M.; Brachmann, R.K.; Haussler, D. Species-specific endogenous retroviruses shape the transcriptional network of the human tumor suppressor protein p53. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 18613–18618. [[CrossRef](#)] [[PubMed](#)]
350. Wang, J.; Bowen, N.J.; Mariño-Ramírez, L.; Jordan, I.K. A c-Myc regulatory subnetwork from human transposable element sequences. *Mol. Biosyst.* **2009**, *5*, 1831–1839. [[CrossRef](#)] [[PubMed](#)]

351. Kunarso, G.; Chia, N.Y.; Jeyakani, J.; Hwang, C.; Lu, X.; Chan, Y.S.; Ng, H.H.; Bourque, G. Transposable elements have rewired the core regulatory network of human embryonic stem cells. *Nat. Genet.* **2010**, *42*, 631–634. [[CrossRef](#)] [[PubMed](#)]
352. Xie, D.; Chen, C.C.; Ptaszek, L.M.; Xiao, S.; Cao, X.; Fang, F.; Ng, H.H.; Lewin, H.A.; Cowan, C.; Zhong, S. Rewirable gene regulatory networks in the preimplantation embryonic development of three mammalian species. *Genome Res.* **2010**, *20*, 804–815. [[CrossRef](#)] [[PubMed](#)]
353. Feschotte, C. Transposable elements and the evolution of regulatory networks. *Nat. Rev. Genet.* **2008**, *9*, 397–405. [[CrossRef](#)] [[PubMed](#)]
354. David, L.; Stolovicki, E.; Haziz, E.; Braun, E. Inherited adaptation of genome-rewired cells in response to a challenging environment. *HFSP J.* **2010**, *4*, 131–141. [[CrossRef](#)] [[PubMed](#)]
355. Scannell, D.R.; Wolfe, K. Rewiring the transcriptional regulatory circuits of cells. *Genome Biol.* **2004**, *5*. [[CrossRef](#)] [[PubMed](#)]
356. Shou, C.; Bhardwaj, N.; Lam, H.Y.; Yan, K.K.; Kim, P.M.; Snyder, M.; Gerstein, M.B. Measuring the evolutionary rewiring of biological networks. *PLoS Comput. Biol.* **2011**, *7*, e1001050. [[CrossRef](#)] [[PubMed](#)]
357. Lowe, C.B.; Kellis, M.; Siepel, A.; Raney, B.J.; Clamp, M.; Salama, S.R.; Kingsley, D.M.; Lindblad-Toh, K.; Haussler, D. Three periods of regulatory innovation during vertebrate evolution. *Science* **2011**, *333*, 1019–1024. [[CrossRef](#)] [[PubMed](#)]
358. Jurka, J.; Bao, W.; Kojima, K.K.; Kohany, O.; Yurka, M.G. Distinct groups of repetitive families preserved in mammals correspond to different periods of regulatory innovations in vertebrates. *Biol. Direct* **2012**, *7*. [[CrossRef](#)] [[PubMed](#)]
359. Huda, A.; Mariño-Ramírez, L.; Landsman, D.; Jordan, I.K. Repetitive DNA elements, nucleosome binding and human gene expression. *Gene* **2009**, *436*, 12–22. [[CrossRef](#)] [[PubMed](#)]
360. Jordan, I.K.; Rogozin, I.B.; Glazko, G.V.; Koonin, E.V. Origin of a substantial fraction of human regulatory sequences from transposable elements. *Trends Genet.* **2003**, *19*, 68–72. [[CrossRef](#)]
361. Jurka, J.; Kapitonov, V.V.; Kohany, O.; Jurka, M.V. Repetitive sequences in complex genomes: Structure and evolution. *Ann. Rev. Genom. Hum. Genet.* **2007**, *8*, 241–259. [[CrossRef](#)] [[PubMed](#)]
362. Jurka, J.; Bao, W.; Kojima, K.K. Families of transposable elements, population structure and the origin of species. *Biol. Direct* **2011**, *6*. [[CrossRef](#)] [[PubMed](#)]
363. Sternberg, R.V.; Shapiro, J.A. How repeated retroelements format genome function. *Cytogenet. Genome Res.* **2005**, *110*, 108–116. [[CrossRef](#)] [[PubMed](#)]
364. Gorbunova, V.; Boeke, J.D.; Helfand, S.L.; Sedivy, J.M. Human genomics. *Sleeping dogs of the genome.* *Science* **2014**, *346*, 1187–1188. [[PubMed](#)]
365. Jern, P.; Coffin, J.M. Effects of retroviruses on host genome function. *Ann. Rev. Genet.* **2008**, *42*, 709–732. [[CrossRef](#)] [[PubMed](#)]
366. Cornelis, G.; Heidmann, O.; Degrelle, S.A.; Vernochet, C.; Lavialle, C.; Letzelter, C.; Bernard-Stoecklin, S.; Hassanin, A.; Mulot, B.; Guillomot, M.; *et al.* Captured retroviral envelope syncytin gene associated with the unique placental structure of higher ruminants. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, E828–E837. [[CrossRef](#)] [[PubMed](#)]
367. Lavialle, C.; Cornelis, G.; Dupressoir, A.; Esnault, C.; Heidmann, O.; Vernochet, C.; Heidmann, T. Paleovirology of “syncytins”, retroviral env genes exapted for a role in placentation. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2013**, *368*. [[CrossRef](#)] [[PubMed](#)]
368. Dupressoir, A.; Lavialle, C.; Heidmann, T. From ancestral infectious retroviruses to bona fide cellular genes: Role of the captured syncytins in placentation. *Placenta* **2012**, *33*, 663–671. [[CrossRef](#)] [[PubMed](#)]
369. Esnault, C.; Priet, S.; Ribet, D.; Vernochet, C.; Bruls, T.; Lavialle, C.; Weissenbach, J.; Heidmann, T. A placenta-specific receptor for the fusogenic, endogenous retrovirus-derived, human syncytin-2. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17532–17537. [[CrossRef](#)] [[PubMed](#)]
370. Chuong, E.B. Retroviruses facilitate the rapid evolution of the mammalian placenta. *Bioessays* **2013**, *35*, 853–861. [[CrossRef](#)] [[PubMed](#)]
371. Macaulay, E.C.; Weeks, R.J.; Andrews, S.; Morison, I.M. Hypomethylation of functional retrotransposon-derived genes in the human placenta. *Mamm. Genome* **2011**, *22*, 722–735. [[CrossRef](#)] [[PubMed](#)]
372. Macaulay, E.C.; Roberts, H.E.; Cheng, X.; Jeffs, A.R.; Baguley, B.C.; Morison, I.M. Retrotransposon hypomethylation in melanoma and expression of a placenta-specific gene. *PLoS ONE* **2014**, *9*, e95840. [[CrossRef](#)] [[PubMed](#)]

373. Renfree, M.B.; Suzuki, S.; Kaneko-Ishino, T. The origin and evolution of genomic imprinting and viviparity in mammals. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2013**, *368*. [[CrossRef](#)] [[PubMed](#)]
374. Lynch, V.J.; Nnamani, M.C.; Kapusta, A.; Brayer, K.; Plaza, S.L.; Mazur, E.C.; Emera, D.; Sheikh, S.Z.; Grützner, F.; Bauersachs, S.; *et al.* Ancient transposable elements transformed the uterine regulatory landscape and transcriptome during the evolution of mammalian pregnancy. *Cell Rep.* **2015**, *10*. [[CrossRef](#)] [[PubMed](#)]
375. Kapusta, A.; Kronenberg, Z.; Lynch, V.J.; Zhuo, X.; Ramsay, L.; Bourque, G.; Yandell, M.; Feschotte, C. Transposable elements are major contributors to the origin, diversification, and regulation of vertebrate long noncoding RNAs. *PLoS Genet.* **2013**, *9*, e1003470. [[CrossRef](#)] [[PubMed](#)]
376. Lynch, V.J.; Leclerc, R.D.; May, G.; Wagner, G.P. Transposon-mediated rewiring of gene regulatory networks contributed to the evolution of pregnancy in mammals. *Nat. Genet.* **2011**, *43*, 1154–1159. [[CrossRef](#)] [[PubMed](#)]
377. Emera, D.; Wagner, G.P. Transposable element recruitments in the mammalian placenta: Impacts and mechanisms. *Briefings Funct. Genom.* **2012**, *11*, 267–276. [[CrossRef](#)] [[PubMed](#)]
378. Emera, D.; Casola, C.; Lynch, V.J.; Wildman, D.E.; Agnew, D.; Wagner, G.P. Convergent evolution of endometrial prolactin expression in primates, mice, and elephants through the independent recruitment of transposable elements. *Mol. Biol. Evol.* **2012**, *29*, 239–247. [[CrossRef](#)] [[PubMed](#)]
379. Goke, J.; Lu, X.; Chan, Y.S.; Ng, H.H.; Ly, L.H.; Sachs, F.; Szczerbinska, I. Dynamic transcription of distinct classes of endogenous retroviral elements marks specific populations of early human embryonic cells. *Cell Stem Cell* **2015**, *16*, 135–141. [[CrossRef](#)] [[PubMed](#)]
380. Grow, E.J.; Flynn, R.A.; Chavez, S.L.; Bayless, N.L.; Wossidlo, M.; Wesche, D.J.; Martin, L.; Ware, C.B.; Blish, C.A.; Chang, H.Y.; *et al.* Intrinsic retroviral reactivation in human preimplantation embryos and pluripotent cells. *Nature* **2015**, *522*, 221–225. [[CrossRef](#)] [[PubMed](#)]
381. Kigami, D.; Minami, N.; Takayama, H.; Imai, H. MuERV-L is one of the earliest transcribed genes in mouse one-cell embryos. *Biol. Reprod.* **2003**, *68*, 651–654. [[CrossRef](#)] [[PubMed](#)]
382. Hutchins, A.P.; Pei, D. Transposable elements at the center of the crossroads between embryogenesis, embryonic stem cells, reprogramming, and long non-coding RNAs. *Sci. Bull.* **2015**, *60*, 1722–1733. [[CrossRef](#)] [[PubMed](#)]
383. Loewer, S.; Cabili, M.N.; Guttman, M.; Loh, Y.H.; Thomas, K.; Park, I.H.; Garber, M.; Curran, M.; Onder, T.; Agarwal, S.; *et al.* Large intergenic non-coding RNA-RoR modulates reprogramming of human induced pluripotent stem cells. *Nat. Genet.* **2010**, *42*, 1113–1117. [[CrossRef](#)] [[PubMed](#)]
384. Kelley, D.; Rinn, J. Transposable elements reveal a stem cell-specific class of long noncoding RNAs. *Genome Biol.* **2012**, *13*. [[CrossRef](#)] [[PubMed](#)]
385. Ng, S.Y.; Stanton, L.W. Long non-coding RNAs in stem cell pluripotency. *Wiley Interdiscip. Rev. RNA* **2013**, *4*, 121–128. [[CrossRef](#)] [[PubMed](#)]
386. Narva, E.; Rahkonen, N.; Emani, M.R.; Lund, R.; Pursiheimo, J.P.; Nästi, J.; Autio, R.; Rasool, O.; Denessiouk, K.; Lähdesmäki, H.; *et al.* RNA-binding protein L1TD1 interacts with LIN28 via RNA and is required for human embryonic stem cell self-renewal and cancer cell proliferation. *Stem Cells* **2012**, *30*, 452–460. [[CrossRef](#)] [[PubMed](#)]
387. Glinsky, G.V. Transposable elements and DNA methylation create in embryonic stem cells human-specific regulatory sequences associated with distal enhancers and noncoding RNAs. *Genome Biol. Evol.* **2015**, *7*, 1432–1454. [[CrossRef](#)] [[PubMed](#)]
388. Kim, D.H.; Marinov, G.K.; Pepke, S.; Singer, Z.S.; He, P.; Williams, B.; Schroth, G.P.; Elowitz, M.B.; Wold, B.J. Single-cell transcriptome analysis reveals dynamic changes in lncRNA expression during reprogramming. *Cell Stem Cell* **2015**, *16*, 88–101. [[CrossRef](#)] [[PubMed](#)]
389. Fort, A.; Hashimoto, K.; Yamada, D.; Salimullah, M.; Keya, C.A.; Saxena, A.; Bonetti, A.; Voineagu, I.; Bertin, N.; Kratz, A.; *et al.* Deep transcriptome profiling of mammalian stem cells supports a regulatory role for retrotransposons in pluripotency maintenance. *Nat. Genet.* **2014**, *46*, 558–566. [[CrossRef](#)] [[PubMed](#)]
390. Schlesinger, S.; Goff, S.P. Retroviral transcriptional regulation and embryonic stem cells: War and peace. *Mol. Cell. Biol.* **2015**, *35*, 770–777. [[CrossRef](#)] [[PubMed](#)]
391. Huo, J.S.; Zambidis, E.T. Pivots of pluripotency: The roles of non-coding RNA in regulating embryonic and induced pluripotent stem cells. *Biochim. Biophys. Acta* **2013**, *1830*, 2385–2394. [[CrossRef](#)] [[PubMed](#)]
392. Hadjiargyrou, M.; Delihias, N. The intertwining of transposable elements and non-coding RNAs. *Int. J. Mol. Sci.* **2013**, *14*, 13307–13328. [[CrossRef](#)] [[PubMed](#)]

393. Kapusta, A.; Feschotte, C. Volatile evolution of long noncoding RNA repertoires: Mechanisms and biological implications. *Trends Genet.* **2014**, *30*, 439–452. [[CrossRef](#)] [[PubMed](#)]
394. Johnson, R.; Guigo, R. The RIDL hypothesis: Transposable elements as functional domains of long noncoding RNAs. *RNA* **2014**, *20*, 959–976. [[CrossRef](#)] [[PubMed](#)]
395. Guttman, M.; Rinn, J.L. Modular regulatory principles of large non-coding RNAs. *Nature* **2012**, *482*, 339–346. [[CrossRef](#)] [[PubMed](#)]
396. Roberts, J.T.; Cardin, S.E.; Borchert, G.M. Burgeoning evidence indicates that microRNAs were initially formed from transposable element sequences. *Mob. Genet. Elem.* **2014**, *4*, e29255. [[CrossRef](#)] [[PubMed](#)]
397. Gifford, W.D.; Pfaff, S.L.; Macfarlan, T.S. Transposable elements as genetic regulatory substrates in early development. *Trends Cell Biol.* **2013**, *23*, 218–226. [[CrossRef](#)] [[PubMed](#)]
398. Piriyaongsa, J.; Marino-Ramirez, L.; Jordan, I.K. Origin and evolution of human microRNAs from transposable elements. *Genetics* **2007**, *176*, 1323–1337. [[CrossRef](#)] [[PubMed](#)]
399. Piriyaongsa, J.; Jordan, I.K. Dual coding of siRNAs and miRNAs by plant transposable elements. *RNA* **2008**, *14*, 814–821. [[CrossRef](#)] [[PubMed](#)]



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